

# The Role of Clusterin, Complement Receptor 1, and Phosphatidylinositol Binding Clathrin Assembly Protein in Alzheimer Disease Risk and Cerebrospinal Fluid Biomarker Levels

Brit-Maren M. Schjeide, BS; Cathrin Schnack, PhD; Jean-Charles Lambert, PhD; Christina M. Lill, MD; Julia Kirchheiner, MD, PhD; Hayrettin Tumani, MD; Markus Otto, MD; Rudolph E. Tanzi, PhD; Hans Lehrach, PhD; Philippe Amouyel, PhD; Christine A. F. von Arnim, MD; Lars Bertram, MD

**Context:** Two recent and simultaneously published genome-wide association studies independently implicated clusterin (*CLU*), complement receptor 1 (*CRI*), and phosphatidylinositol binding clathrin assembly protein (*PICALM*) as putative novel Alzheimer disease (AD) risk loci. Despite their strong statistical support, all 3 signals emerged from heterogeneous case-control populations and lack replication in different settings.

**Objective:** To determine whether genetic variants in *CLU*, *CRI*, and *PICALM* confer risk for AD in independent data sets ( $n=4254$ ) and to test the impact of these markers on cerebrospinal fluid (CSF)-A $\beta$ 42 and total-tau protein levels ( $n=425$ ).

**Design:** Genetic association study using family-based and case-control designs.

**Setting:** Ambulatory or hospitalized care.

**Participants:** Family samples originate from mostly multiplex pedigrees recruited at different centers in the United States (1245 families, 2654 individuals with AD, and 1175 unaffected relatives). Unrelated case-control subjects originate from 1 clinical center in Germany (214 individuals with AD and 211 controls). All subjects were of European descent.

**Main Outcome Measures:** The association between 5 genetic variants in *CLU*, *CRI*, and *PICALM* and risk for AD, and the correlation between these 5 genetic variants and CSF-A $\beta$ 42 and tau levels.

**Results:** All 3 investigated loci showed significant associations between risk for AD (1-tailed  $P$  values ranging from  $<.001$  to  $.02$ ) and consistent effect sizes and direction. For each locus, the overall evidence of association was substantially strengthened on meta-analysis of all available data (2-tailed  $P$  values ranging from  $1.1 \times 10^{-16}$  to  $4.1 \times 10^{-7}$ ). Of all markers tested, only rs541458 in *PICALM* was shown to have an effect on CSF protein levels, suggesting that the AD risk allele is associated with decreased CSF A $\beta$ 42 levels (2-tailed  $P=.002$ ).

**Conclusions:** This study provides compelling independent evidence that genetic variants in *CLU*, *CRI*, and *PICALM* are genetically associated with risk for AD. Furthermore, the CSF biomarker analyses provide a first insight into the potentially predominant pathogenetic mechanism(s) underlying the association between AD risk and *PICALM*.

*Arch Gen Psychiatry.* 2011;68(2):207-213

A LARGE PROPORTION OF SUSCEPTIBILITY TO non-Mendelian (sporadic or late-onset) forms of Alzheimer disease (AD) is likely determined by the contribution of common genetic risk factors that exert their effects with low penetrance.<sup>1</sup> Besides a well-established and highly significant association between risk for AD and genetic variants in the apolipoprotein E (*APOE*) gene on chromosome 19q, the exact number and nature of the remaining susceptibility loci remain elusive.<sup>2</sup> More than 600 candidate loci have been tested in mostly hypothesis-driven genetic association studies in nearly 1500 publications, but only a few

of these showed significant effects in systematic meta-analyses<sup>3</sup> (for up-to-date results, see the AlzGene database at <http://www.alzgene.org> maintained by our group the Alzheimer Research Forum). High-throughput genotyping technologies now enable researchers to perform genome-wide analyses in an unbiased and largely hypothesis-free fashion, using sets of densely spaced single-nucleotide polymorphisms (SNPs). Within the past 3 years, 13 such genome-wide association studies (GWASs) have been published in the field of AD, highlighting more than 30 novel potential susceptibility loci with essentially no overlap in results across studies with the exception of *APOE*.<sup>4</sup> The 2

Author Affiliations are listed at the end of this article.

most recent GWASs implicated SNPs in clusterin (*CLU*), complement receptor 1 (*CR1*), and phosphatidylinositol binding clathrin assembly protein (*PICALM*) as novel putative AD risk loci.<sup>5,6</sup> Although the association with *CLU* independently reached genome-wide significance in both GWASs, the genetic markers in *CR1* and *PICALM* were each initially identified in only 1 of the 2 studies but were subsequently replicated at subgenome-wide significance in the other. In systematic meta-analyses of all genetic association data available on AD, these associations are among the highest ranking findings of the entire field. Notwithstanding their relatively strong statistical support, all 3 signals emerged from heterogeneous multicenter case-control studies, which still lack replication by independent groups, especially in samples ascertained from families with AD. The purpose of our study was to assess the role of *CLU*, *CR1*, and *PICALM* in AD risk in a collection of more than 4250 white subjects originating from multiplex families with AD ascertained in the United States and in unrelated AD cases and controls recruited in Germany. In addition, all 3 loci were tested for their potential link to cerebrospinal fluid (CSF) levels of 2 established AD biomarkers in the German data set ( $n=425$ ). Finally, all results were combined with the previously published evidence via random-effects meta-analysis and were classified on the basis of their epidemiological credibility.

## METHODS

### STUDY SUBJECTS

All samples were collected with informed written consent and appropriate ethical approval at the collection sites. Demographic details of the various data sets can be found in eTable 1 (available at <http://www.archgenpsychiatry.com>). The family-based samples were ascertained in the United States and originate from 4 different projects aimed at the study of genetic factors in AD: the Consortium on Alzheimer's Genetics sample,<sup>7</sup> the National Institute of Mental Health sample,<sup>8</sup> and the National Institute on Aging sample and the National Cell Repository for Alzheimer's Disease sample (both at <http://www.ncrad.org>). With the exception of the Consortium on Alzheimer's Genetics sample, the majority of pedigrees analyzed herein were nuclear families ascertained on the basis of multiple affected individuals, generally lacking parental genotypes, because parents were usually deceased at the time of proband recruitment. In addition to containing at least 1 affected relative pair, many pedigrees also had DNA available from additional affected or unaffected individuals (mostly siblings). The diagnosis of definite, probable, or possible AD was made according to criteria from the National Institute of Neurological and Communicative Diseases and Stroke and the Alzheimer's Disease and Related Disorders Association for affected individuals in all 4 samples.<sup>9</sup> Only families of self-reported white ancestry in which no affected individual showed an onset at age younger than 50 years were included.

The case-control data set was obtained from individuals recruited between 1999 and 2008 at the Memory Clinic of the Neurology University Hospital in Ulm, Germany. Alzheimer disease was diagnosed according to the National Institute of Neurological and Communicative Diseases and Stroke–Alzheimer's Disease and Related Disorders Association criteria<sup>9</sup> and the *DSM-IV*. Only patients fulfilling diagnostic crite-

ria for probable AD were included. Unrelated control subjects were recruited at the same site and did not display any cognitive or neurological deficits following thorough clinical (including magnetic resonance imaging or computed tomography) and neuropsychological examination. Although the presence of population admixture within this data set could not be assessed owing to a lack of sufficient genotype information, it appears unlikely to be an issue given that both cases and controls were collected in a relatively homogenous area from southern Germany.

### GENOTYPING

All genetic analyses were performed at the Max Planck Institute for Molecular Genetics in Berlin, Germany, and were performed blind to the clinical status of the subjects. Genotyping of SNPs was performed in 384-well format using TaqMan chemistry (Applied Biosystems, Carlsbad, California) according to manufacturer's instructions. SNP rs541458 in *PICALM* was genotyped using a Singleplex TaqMan genotyping assay (Applied Biosystems, Carlsbad, California), whereas the other 4 SNPs (rs11136000, rs2279590, and rs9331888 in *CLU* and rs6656401 in *CR1*) were genotyped in parallel on the OpenArray (Applied Biosystems) multiplex genotyping system. Overall, genotyping efficiency was greater than 98%, with an error rate of less than 0.2% (based on more than 1500 genotypes generated across HapMap samples run in multiples on each genotyping plate). *APOE* genotypes in the German case-control cohort were determined by direct sequencing on an ABI3730XL genetic analyzer (Applied Biosystems) or from blood samples via isoelectric focusing. None of the markers violated the Hardy-Weinberg equilibrium at  $P \leq .05$  in the control samples.

### MEASUREMENTS OF CSF BIOMARKERS

Cerebrospinal fluid biomarker data were available only for the German case-control sample. The CSF sample collection and preanalytic processing were performed at the Neurology University Hospital in Ulm, Germany, using a standardized protocol as previously described elsewhere.<sup>10</sup> In brief, CSF samples were placed into polypropylene tubes following lumbar puncture, centrifuged immediately after collection, and stored within 2 hours at  $-80^{\circ}\text{C}$  in Eppendorf tubes until analysis. CSF-A $\beta$ 42 and total tau protein levels were determined using commercially available sandwich enzyme-linked immunosorbent assays Innostest  $\beta$ -amyloid[1-42] and hTau Ag kits; Innogenetics, Gent, Belgium following the manufacturers instructions as previously described elsewhere.<sup>11,12</sup> Monitoring the diagnostic accuracy of the CSF tests was done according to international guidelines.<sup>13</sup>

### STATISTICAL ANALYSIS

Genetic association analyses using affection status as a dichotomous trait were performed assuming additive transmission models using FBAT version 2.0.3 (Harvard School of Public Health, Cambridge, Massachusetts; <http://www.biostat.harvard.edu/~lbat/default.html>) for the family-based sample (with an equal-weight offset correction [ie, identical weights of opposite sign are assigned to affected and unaffected individuals resulting in a statistic that contrasts transmissions to affected vs unaffected individuals], applying the empirical variance estimation function to account for the presence of multiple affected individuals per nuclear family)<sup>14</sup> and PLINK version 1.07 (<http://pngu.mgh.harvard.edu/~purcell/plink/>) for the case-control sample. To increase power, genotypes for all 4 fam-

ily-based data sets were pooled before analysis. Odds ratios (ORs) and confidence intervals (CIs) in the case-control sample were calculated using PLINK, whereas ORs and CIs in the family-based sample were determined by conditional logistic regression stratified by family using SAS version 9.2 (SAS Institute, Cary, North Carolina) as described previously elsewhere.<sup>15</sup> Combined evidence of association across the family-based and case-control data sets was determined using random effects via the *rmeta* package version 2.16 in R version 2.10.0 (R Foundation for Statistical Computing, Vienna, Austria). Violations in the Hardy-Weinberg equilibrium in unaffected individuals (determined for all German controls and separately for a collection of unaffected individuals from the US family samples [1 per family, where available]) were determined using PLINK. Because the hypothesis of this part of our study was to specifically probe for the previously reported allele-specific effects resulting in increased (rs6656401 and rs9331888) or decreased (rs11136000, rs2279590, and rs541458) ORs in carriers of the minor vs major alleles at these sites, statistical significance is expressed as 1-tailed *P* values and 90% CIs for all the outlined analyses.

Genetic association analyses using CSF biomarker levels as quantitative traits were calculated using PLINK via linear regression, including age, sex, and *APOE* ε4 dose (coded as 0,1,2) as covariates. To approximate a normal distribution, both CSF-Aβ42 and total-tau concentrations were log-transformed (base 10) before analysis. The quantitative trait analyses were then performed for all individuals combined (including affection status as a covariate) as well as for AD cases and healthy controls separately. Meta-analyses combining the genotype data generated in our study with data from previously published association studies on the same SNPs<sup>5,6,16-18</sup> were based on random-effect models<sup>19</sup> using crude ORs and standard errors calculated for each data set. For the assessment of the epidemiologic credibility of these meta-analyses, we also performed sensitivity analyses after exclusion of data sets in which control subjects had genotypes that violated the Hardy-Weinberg equilibrium and after exclusion of the initial study. Between-study heterogeneity was quantified using the *I*<sup>2</sup> metric. Evidence for reporting bias was assessed using a modified regression test.<sup>20</sup> Statistical significance for all meta-analyses was calculated as 2-tailed *P* values and 95% CIs. Power analysis for the case-control sample was performed using the Genetic Power Calculator (<http://pngu.mgh.harvard.edu/~purcell/gpc/>); power analysis for the family-based samples was estimated using PBAT version 3.6 (<http://www.biostat.harvard.edu/~clange/default.htm>). Both analyses were based on a disease prevalence of 0.1 and a 1-sided  $\alpha$  level of .05, which were based on allelic ORs reported for each SNP in the AlzGene database. Power calculations of the meta-analyses combining our data with previously generated data were based on a 2-sided genome-wide  $\alpha$  level of  $5 \times 10^{-8}$ .

#### ASSESSMENT OF EPIDEMIOLOGIC CREDIBILITY OF META-ANALYSIS RESULTS

All meta-analysis results were graded on the basis of the Human Genome Epidemiology Network interim criteria for the assessment of cumulative evidence of genetic associations, referred to as the Venice criteria.<sup>21,22</sup> These criteria take into account the amount of evidence (ie, sample size as measured by the number of minor alleles across both cases and controls), the consistency of replication (ie, heterogeneity across studies measured by the *I*<sup>2</sup> statistic), and protection from bias (ie, via the modified regression test and sensitivity analyses, excluding the initial study and samples showing Hardy-Weinberg equilibrium violations at  $P \leq .05$  in controls). For more details about how these criteria are applied, see Ioannidis et al,<sup>21</sup> Khoury et al,<sup>22</sup> Allen et al,<sup>23</sup> and <http://www.alzgene.org/methods.asp>.

Association analyses of the family-based samples revealed nominal association (based on 1-tailed *P* values) with 3 of the 5 tested SNPs (one each in *CLU*, *CRI*, *PICALM*; **Table 1**); a fourth SNP in *CLU* showed a statistical trend in favor of association (rs11136000; 1-tailed  $P = .06$ ). Effect sizes and effect directions were consistent with those reported in the original GWASs for each of these associations (Table 1, eTable 2, and eFigures 1, 2, and 3). In the case-control sample, nominally significant associations (based on 1-tailed *P* values) were observed for 2 of the 5 tested SNPs (one each in *CRI* and *PICALM*; Table 1). Here, too, the underlying effects were very consistent with both the family-based findings and the previous GWAS results.<sup>5,6</sup> The only gene not showing evidence for association in the case-control sample was *CLU* (best SNP rs9331888; 1-tailed  $P \sim .12$ ), which also showed the weakest support for association in the family-based samples. However, assuming an allelic OR of approximately 1.15, the power to detect a nominal, 1-tailed *P* value of .05 in the case-control sample alone was only approximately 30% (compared with ~60% in the family sample and ~70% in the combined sample), suggesting that the lack of significance for *CLU* in the case-control data set could be due to a lack of power. Combining the results from the family-based and case-control data sets of our study via random-effects meta-analysis increased the statistical support for all 3 loci (Table 1), with *CRI* showing the most pronounced and most significant effects (OR, 1.33; 1-tailed  $P < .001$ ) and *CLU* showing the smallest change in AD risk (OR, 0.88; 1-tailed  $P \sim .04$ ). Finally, meta-analyses of our data and the data from the previous studies yielded highly significant associations for all 3 loci. In these analyses, the rs11136000 variant in *CLU* showed by far the strongest support for association (OR, 0.86; 2-tailed  $P = 1.1 \times 10^{-16}$ ), followed by *PICALM* (OR, 0.87; 2-tailed  $P = 2.3 \times 10^{-11}$ ) and *CRI* (OR, 1.21; 2-tailed  $P = 4.1 \times 10^{-7}$ ). Application of the Human Genome Epidemiology Network criteria for the cumulative assessment of genetic association studies assigned “strong” epidemiologic credibility to the *CLU* and *PICALM* findings (1 SNP each) and “moderate” credibility to the *CRI* finding. The latter can be attributed to heterogeneity across the GWAS follow-up data by Lambert et al,<sup>6</sup> in particular the stage 2 case-control sample from Italy, which shows a slightly opposite direction of effect when compared with the rest of the samples from all of the studies, resulting in an *I*<sup>2</sup> of 36% (eFigure 2). Removal of that 1 outlying sample considerably strengthened the overall evidence in favor of *CRI* (OR, 1.25; 2-tailed  $P = 4.1 \times 10^{-14}$ ; strong epidemiological credibility with an *I*<sup>2</sup> of 0%).

Next, we investigated whether or not any of the 5 investigated SNPs in *CLU*, *CRI*, and *PICALM* showed evidence for association with levels of CSF-Aβ42 and total tau, 2 well-established biomarkers for AD.<sup>24</sup> All analyses were performed in the combined case-control sample, as well as in AD cases and controls separately. As can be seen in **Table 2**, the *PICALM* T allele (associated with AD risk) showed a significant association with de-

**Table 1. Association Analyses of Clusterin, Complement Receptor 1, and Phosphatidylinositol Binding Clathrin Assembly Protein<sup>a</sup>**

Chromosome	Gene	SNP	Study Sample	MAF	P Value	OR (CI)	Power	Credibility
1	<i>CR1</i>	rs6656401	United States	0.203	.027	1.28 (1.08-1.51)	0.53	
			Germany	0.190	<.001	1.49 (1.12-1.97)	0.29	
			Combined	0.200	.001	1.33 (1.15-1.54)	0.67	
			Meta-analysis		$4.1 \times 10^{-7}$	1.20 (1.12-1.30)	0.94	
8	<i>CLU</i>	rs11136000	United States	0.377	.06	0.84 (0.73-0.96)	0.60	Moderate
			Germany	0.378	.49	0.99 (0.83-1.27)	0.32	
			Combined	0.377	.04	0.88 (0.78-0.99)	0.72	
			Meta-analysis		$1.1 \times 10^{-16}$	0.86 (0.83-0.89)	0.99	
8	<i>CLU</i>	rs2279590	United States	0.393	.03	0.83 (0.73-0.95)	0.61	Strong
			Germany	0.402	.63	1.05 (0.83-1.33)	0.32	
			Combined	0.395	.38	0.91 (0.73-1.13)	0.73	
			Meta-analysis		$5.1 \times 10^{-10}$	0.86 (0.82-0.90)	0.90	
8	<i>CLU</i>	rs9331888	United States	0.298	.19	1.06 (0.92-1.23)	0.48	Weak
			Germany	0.279	.12	1.20 (0.93-1.55)	0.29	
			Combined	0.294	.12	1.10 (0.97-1.24)	0.67	
			Meta-analysis		$2.7 \times 10^{-7}$	1.14 (1.09-1.20)	0.71	
11	<i>PICALM</i>	rs541458	United States	0.299	.02	0.87 (0.75-1.02)	0.49	
			Germany	0.358	.02	0.72 (0.56-0.93)	0.26	
			Combined	0.312	.01	0.82 (0.71-0.95)	0.59	
			Meta-analysis		$2.3 \times 10^{-11}$	0.87 (0.83-0.90)	0.91	

Abbreviations: CI, confidence interval; *CLU*, clusterin; *CR1*, complement receptor 1; MAF, minor allele frequency; OR, odds ratio; *PICALM*, phosphatidylinositol binding clathrin assembly protein; SNP, single-nucleotide polymorphism.

<sup>a</sup>Family-based association analyses (our US sample) were performed using FBAT version 2.0.3, and case-control analyses (our German sample) were performed using PLINK version 1.07. Results from both analyses were combined using random-effects models. *P* values for all analyses are 1-tailed with 90% CIs, except for the meta-analysis results and for marker rs2279590 in *CLU*, for which a 2-tailed *P* value and 95% CIs are reported owing to the opposite direction of effect across the family-based and case-control data sets. Meta-analyses were performed by combining sample-specific ORs of the US and German samples with those calculated from publicly available genotype data in other samples, as listed on the AlzGene database. Note that individuals with unknown diagnoses were not used in any of the statistical analyses. Power estimates are for 1-sided *P* values of .05, except for the overall meta-analyses in which power was estimated for 2-sided genome-wide significance ( $5 \times 10^{-8}$ ). Credibility refers to the assessment of the overall epidemiological credibility of the meta-analysis results using the Human Genome Epidemiology Network interim guidelines.

creased CSF-A $\beta$ 42 levels (2-tailed *P* = .002; **Figure 1**). The decrease in CSF-A $\beta$ 42 levels was dependent on dose, with the strongest effect in homozygous carriers of the T allele with an approximately 20% decrease in absolute A $\beta$ 42 concentration when compared with homozygous carriers of the C allele. Although the direction of this association is consistent with that on the same trait conferred by the *APOE*  $\epsilon$ 4 allele, it is less pronounced and much less significant (~50% decrease in  $\epsilon$ 4/4 vs  $\epsilon$ 3/3; 2-tailed *P* =  $7.2 \times 10^{-16}$ ; Table 2 and eFigure 4). Furthermore, as has been described in previous studies (eg, Kester et al<sup>25</sup>), the *APOE* effect on CSF-A $\beta$ 42 levels was most evident in unaffected individuals (2-tailed *P* =  $2.4 \times 10^{-11}$ ), whereas the association for *PICALM* was strongest in the AD case group (2-tailed *P* = .01). For CSF-A $\beta$ 42 levels, the only other marker that approached statistical significance was rs6656401 in *CR1* (2-tailed *P* = .08, in controls only). Owing to the only weak statistical support and heterogeneity of the effect across genotypes (Table 2), this likely represents a chance finding. Finally, apart from *APOE* (2-tailed *P* ~ .003), no association was observed between any of the tested GWAS SNPs and levels of CSF total tau (Table 2).

#### COMMENT

Even in the GWAS era, independent replication remains the primary means of distinguishing true-positive vs false-positive genetic association findings.<sup>26,27</sup> Herein, we provide compelling evidence that all

3 of the recently proposed novel AD candidate loci, *CLU*, *CR1*, and *PICALM*, show association with risk for AD in a study of more than 4250 subjects from the United States and Germany. For the lead signals in all 3 genes, the effect direction and effect sizes estimated were remarkably consistent with what was originally reported. Intriguingly, 90% of our sample was from mostly multiplex AD families and analyzed using family-based methods, an approach that is generally believed to be less prone to bias due to undetected population admixture.<sup>28</sup> Thus, the convergence of independent case-control and family-based findings lends further support to the notion that *CLU*, *CR1*, and *PICALM*, indeed, represent genuine AD susceptibility factors. In addition to the risk effects, we report novel evidence suggesting a link between CSF-A $\beta$ 42 levels and allele dose at the *PICALM* rs541458 SNP, which, to the best of our knowledge, was not described in any previous study. If validated in independent cohorts, this finding could provide a first clue regarding the predominant pathogenetic mechanism underlying the association between AD and *PICALM*.

Although functional data are still lacking to elucidate the precise mechanism by which SNPs in or near *PICALM* could impact levels of A $\beta$  in the brain and CSF of risk-allele carriers, it is tempting to speculate that dysfunction of the *PICALM* protein could be connected to amyloid precursor protein processing via endocytic pathways.<sup>29</sup> This hypothesis was already outlined by Harold et al<sup>5</sup> in their original GWAS and is based on the notion that the *PICALM* protein is involved in clathrin-

**Table 2. Effect of Genome-Wide Association Studies' Loci on Cerebrospinal Fluid Biomarkers<sup>a</sup>**

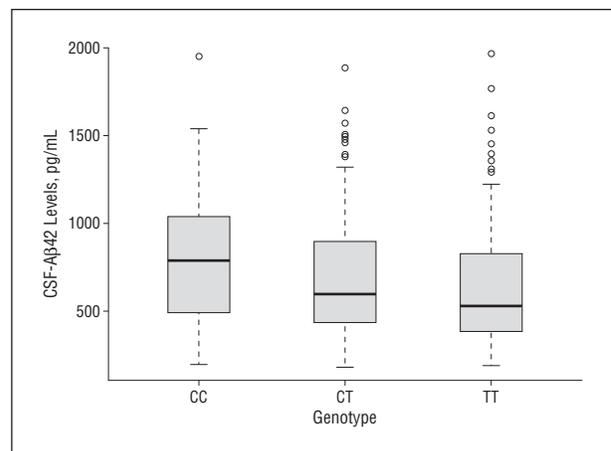
Chromosome, Gene	SNP	CSF Trait	Sample	Mean (SD) CSF Level per Genotype, pg/mL									Wald Test	P Value
				Subjects, No.	Min/Min	Subjects, No.	Min/Maj	Subjects, No.	Maj/Maj	Subjects, No.	Min/Maj	Subjects, No.		
1, <i>CR1</i>	rs6656401	Aβ42	All	416	590.5 (406.6)	22	697.1 (361.9)	137	659.4 (340)	257	-0.2356	.81		
				AD	202	497.5 (355.1)	15	473.5 (184.1)	75	465.9 (202.8)	112	0.157	.88	
			Controls	187	831.5 (494.9)	6	999.4 (314.8)	59	873.2 (326.6)	122	1.793	.08		
				Total tau	416	445 (339.9)	22	470.4 (329.8)	137	451.9 (360.5)	257	0.8548	.39	
			AD	200	534.5 (370.2)	15	629.3 (343.8)	74	664.4 (410)	111	-0.8253	.41		
				Controls	188	262.3 (158.9)	6	270.6 (162)	59	271.3 (178.1)	123	0.3295	.74	
8, <i>CLU</i>	rs11136000	Aβ42	All	413	637.2 (337.9)	59	680.2 (353.1)	194	669.8 (352.7)	160	-0.674	.50		
				AD	198	459.4 (242.6)	27	476 (218.1)	95	470.1 (192.5)	76	-0.2853	.78	
			Controls	187	835.3 (337.1)	27	925.7 (320.5)	90	922.6 (339.5)	70	-1.135	.26		
				Total tau	413	404.4 (278.5)	59	457.5 (349.4)	192	470.3 (368.4)	162	-1.153	.25	
			AD	196	559.9 (269)	27	646.3 (373.7)	93	661.3 (429)	76	-0.6507	.52		
				Controls	188	275.7 (215.2)	27	263.1 (177.4)	90	274.4 (143.8)	71	-0.8586	.39	
11, <i>PICALM</i>	rs541458	Aβ42	All	412	808.7 (393.6)	38	692 (343.5)	193	630.6 (346.1)	181	3.065	.002		
				AD	196	540.1 (259.8)	13	507.7 (235.9)	87	439.7 (182.3)	96	2.601	.01	
			Controls	190	985 (375.5)	23	888.2 (324)	93	912.3 (337.2)	74	0.3272	.74		
				Total tau	413	357.9 (283.7)	37	454.6 (370.7)	193	466.2 (323.3)	183	-1.294	.20	
			AD	195	556.3 (407.2)	12	652.4 (436.3)	87	620 (325.8)	96	-0.186	.85		
				Controls	191	256.4 (124.3)	23	275.9 (174.6)	93	278.7 (184.3)	75	-0.3105	.76	
19, <i>APOE</i>	ε4 vs ε3	Aβ42	All	395	410 (157.2)	52	561.6 (237.1)	152	808.2 (403.7)	191	-8.422	$7.15 \times 10^{-16}$		
				AD	200	400.8 (151.8)	41	486.2 (137.7)	90	498.8 (290.2)	69	-1.866	.06	
			Controls	169	485.4 (183.7)	8	728 (279.6)	47	1020 (330.5)	114	-7.171	$2.4 \times 10^{-11}$		
				Total tau	396	511.7 (324.9)	51	548.5 (352)	153	401.3 (345.7)	192	3.037	.003	
			AD	199	547.2 (322.3)	40	684.4 (346)	90	639.1 (459.9)	69	-0.2284	.82		
				Controls	170	354.2 (276)	8	336.2 (240.3)	47	258.6 (127.5)	115	0.688	.49	

Abbreviations: AD, Alzheimer disease; *APOE*, apolipoprotein E; *CLU*, clusterin; *CR1*, complement receptor 1; CSF, cerebrospinal fluid; Maj/Maj, major allele heterozygotes; Min/Maj, heterozygotes; Min/Min, minor allele homozygotes; *PICALM*, phosphatidylinositol binding clathrin assembly protein; SNP, single-nucleotide polymorphism.

<sup>a</sup>Results are based on the case-control sample from Germany. The *t* statistics and 2-tailed *P* values are based on log-normalized (base 10) CSF-Aβ42 and total-tau protein levels and were computed via linear regression using PLINK version 1.0.7, adjusting for age, sex, and *APOE* ε4 allele (not in the analyses testing *APOE* itself) dose in subjects with AD and in controls, and also diagnostic status in analyses of all individuals. All *P* values are 2-tailed. The corresponding results for *APOE* are displayed here only for comparison. Note that individuals with unknown diagnoses were not used in any of the statistical analyses. See Figure 1 and eFigure 4 for a graphical depiction of the distribution of Aβ42 levels across *PICALM* and *APOE* genotypes, respectively.

mediated endocytosis,<sup>30</sup> the inhibition of which can lead to a reduction in amyloid precursor protein internalization and Aβ production.<sup>31</sup> Therefore, it is conceivable that sequence variants in or close to the *PICALM* gene could impact this process, either directly or via changes in synaptic activity. However, these hypotheses are largely speculative and still need to be addressed in specific molecular genetic and biochemical experiments.

Although our study significantly strengthens the overall evidence implying that all 3 recently proposed GWAS signals represent genuine AD susceptibility loci, a number of questions remain unanswered. First, the genetic markers tested in our study (and in the primary GWAS) are very likely not the functional genetic variants. In fact, none of the 5 variants maps compellingly close to, or is in significant linkage disequilibrium with, any obviously functional variant in these regions.<sup>5,6</sup> Thus, despite the nearly unequivocal evidence suggesting that the genomic regions near the investigated SNPs likely contain functionally active variant(s) relevant in AD pathogenesis, more work is needed to further pinpoint the location and characterize the role of the pathophysiologically active elements. Second, although these genes appear to exert their effects across the majority of the white populations investigated to date, other ancestral backgrounds need to be studied with high priority to arrive at a better understanding of the role that these genes play in other populations. Third, although our CSF biomarker analyses suggest that the AD association with



**Figure 1.** Box plot of the distribution of absolute CSF-Aβ42 levels across the 3 genotypes observed at rs541458 in *PICALM* in the combined case-control data set from Germany. Horizontal lines represent median values, boxes are 25% to 75% ranges, and whiskers extend to  $1 \times$  the interquartile range; values outside this range are depicted as circles. CSF-Aβ42 levels decrease with increasing numbers of T alleles (which are associated with risk of Alzheimer disease). Using log-normalized Aβ42 levels, we found that this effect is significant at  $P = .002$  after adjusting for age, sex, apolipoprotein E (*APOE*) ε4 dose, and diagnostic group. CSF indicates cerebrospinal fluid. A similar, albeit more pronounced and more significant, effect is also observed for increasing numbers of the ε4 allele at the *APOE* locus (eFigure 4).

*PICALM* is functionally more likely related to amyloid precursor protein-Aβ metabolism than to tau dysfunction, it cannot be excluded that this association is actu-

ally the result of another, correlated effect or that it represents a false-positive finding altogether (although the association between *PICALM* and CSF-A $\beta$ 42 levels remains nominally significant even after Bonferroni correction for multiple testing [20 tests] with a 2-tailed  $P \sim .047$ ).

Likewise, our failure to observe significant changes in CSF-A $\beta$ 42 levels with variants in the other 2 genes and the lack of significant effects on CSF total-tau levels of all 3 genes either imply that these effects do not exist (at least not with the markers studied) or that our sample lacked power to detect them. However, in our case-control cohort, assuming a 2-sided  $\alpha$  level of 5% showed that we had more than 80% power to detect additive genetic effects that explain down to 2% of the total variance of A $\beta$ 42 and total-tau CSF levels. Therefore, unless the underlying genetic effects leading to changes in the levels of these biomarkers are even smaller, lack of power is likely not an issue. Finally, despite the overall strong statistical and epidemiological support for the associations between AD risk and *CLU*, *CRI*, and *PICALM*, the relevance of these findings on a population-wide level remains to be determined. Similar to what is found in recent GWASs for other complex diseases,<sup>26,27</sup> the observed risk effects are relatively small; that is, they confer changes in disease risk between approximately 15% and 20% per allele. As a result, the associations described by us and others will likely have no major role serving as a diagnostic or predictive tool for AD in a clinical setting, unless other (eg, rarer) variants of higher penetrance are linked to the observed effects. Nevertheless, depending on their precise functional role in AD pathogenesis, the highlighted loci might still be essential in advancing our understanding of the biochemical processes leading to AD and in developing appropriate and effective means to target these processes therapeutically.

In summary, we provide compelling evidence that genetic variants in *CLU*, *CRI*, and *PICALM* are genetically associated with risk for AD. The independent convergence of case-control GWASs and family-based follow-up data substantially strengthens the notion that these genes (likely in concert with numerous other loci) exert genuine disease-modifying effects. The results from our CSF biomarker analyses suggest that the predominant pathogenetic mechanism(s) underlying the association between AD risk and *PICALM* warrants examination in future functional genetic studies.

**Submitted for Publication:** May 4, 2010; final revision received August 9, 2010; accepted October 5, 2010.

**Author Affiliations:** Department of Vertebrate Genomics, Max Planck Institute for Molecular Genetics, Berlin (Mr Schjeide and Drs Lill, Lehrach, and Bertram), Department of Neurology, University Hospital, Ulm (Drs Schnack, Kirchheiner, Tumani, Otto, and von Arnim), and Department of Neurology, University Medicine, Johannes Gutenberg University, Mainz (Dr Lill), Germany; Institut National de la Santé et de la Recherche Médicale and Institut Pasteur de Lille (Drs Lambert and Amouyel) and Université Lille, Nord de France, Lille, France (Dr Amouyel); and Genetics and Aging Re-

search Unit, Massachusetts General Hospital, Harvard Medical School, Boston (Dr Tanzi).

**Correspondence:** Lars Bertram, MD, Neuropsychiatric Genetics Group, Department Vertebrate Genomics, Max Planck Institute for Molecular Genetics, Ihnestrasse 63, Rm 204.1, 14195 Berlin, Germany (bertram@molgen.mpg.de).

**Author Contributions:** Ms Schjeide and Dr Bertram had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis.

**Financial Disclosure:** None reported.

**Funding/Support:** The project was funded by grants from the Institut Pasteur Lille, the Fondation Plan Alzheimer, the Cure Alzheimer's Fund (Dr Bertram), and the German Federal Ministry of Research and Education (Dr Bertram). Dr von Arnim was supported by the WIN-Kolleg of the Heidelberg Academy of Sciences and Humanities. Dr Otto was supported by Antepriion, cNeuro, NeuroTAS, and Landesstiftung Baden-Württemberg.

**Role of the Sponsor:** The funding sources had no role in the design and conduct of the study; in the collection, analysis, and interpretation of the data; or in the preparation, review, or approval of the manuscript.

**Online-Only Material:** The eFigures and eTables 1 and 3 are available at <http://www.archgenpsychiatry.com>.

**Additional Information:** Samples from the National Cell Repository for Alzheimer's Disease, which receives government support under a cooperative agreement grant (U24 AG21886) awarded by the National Institute on Aging, were used in this study. Since the submission of our manuscript, a number of studies have been published that, like our study, independently confirm the association between AD risk and genetic polymorphisms in *CLU*, *CRI*, and *PICALM*. Please see <http://www.alzgene.org> for up-to-date results.

**Additional Contributions:** We thank all patients and other participants across the various samples for their contribution to this study. We also thank Mrs Sabina Gualazzini for her help with data collection and management of the Ulm samples. We thank the contributors, including the Alzheimer's Disease Centers for their collection of samples that were used in this study, as well as patients and their families, whose help and participation made this work possible.

## REFERENCES

1. Bertram L, Tanzi RE. Thirty years of Alzheimer's disease genetics: the implications of systematic meta-analyses. *Nat Rev Neurosci*. 2008;9(10):768-778.
2. Saunders AM, Strittmatter WJ, Schmechel D, St. George-Hyslop PH, Pericak-Vance MA, Joo SH, Rosi BL, Gusella JF, Crapper-MacLachlan DR, Alberts MJ, Hulette C, Crain B, Goldgaber D, Roses AD. Association of apolipoprotein E allele epsilon 4 with late-onset familial and sporadic Alzheimer's disease. *Neurology*. 1993;43(8):1467-1472.
3. Bertram L, McQueen MB, Mullin K, Blacker D, Tanzi RE. Systematic meta-analyses of Alzheimer disease genetic association studies: the AlzGene database. *Nat Genet*. 2007;39(1):17-23.
4. Bertram L, Tanzi RE. Genome-wide association studies in Alzheimer's disease. *Hum Mol Genet*. 2009;18(R2):R137-R145.
5. Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, Pahwa JS, Moskva V, Dowzell K, Williams A, Jones N, Thomas C, Stretton A, Morgan AR, Lovestone S, Powell J, Proitsi P, Lupton MK, Brayne C, Rubinsztein DC, Gill M, Lawlor B, Lynch A, Morgan K, Brown KS, Passmore PA, Craig D, McGuinness

- B, Todd S, Holmes C, Mann D, Smith AD, Love S, Kehoe PG, Hardy J, Mead S, Fox N, Rossor M, Collinge J, Maier W, Jessen F, Schürmann B, van den Bussche H, Heuser I, Kornhuber J, Wilfang J, Dichgans M, Frölich L, Hampel H, Hüll M, Rujescu D, Goate AM, Kauwe JS, Cruchaga C, Nowotny P, Morris JC, Mayo K, Sleegers K, Bettens K, Engelborghs S, De Deyn PP, Van Broeckhoven C, Livingston G, Bass NJ, Gurling H, McQuillin A, Gwilliam R, Deloukas P, Al-Chalabi A, Shaw CE, Tsolaki M, Singleton AB, Guerreiro R, Mühleisen TW, Nöthen MM, Moebus S, Jöckel KH, Klopp N, Wichmann HE, Carrasquillo MM, Pankratz VS, Younkin SG, Holmans PA, O'Donovan M, Owen MJ, Williams J. Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease [published correction appears in *Nat Genet.* 2009;41(10):1156]. *Nat Genet.* 2009;41(10):1088-1093.
6. Lambert JC, Heath S, Even G, Campion D, Sleegers K, Hiltunen M, Combarros O, Zelenika D, Bullido MJ, Tavernier B, Letenneur L, Bettens K, Berr C, Pasquier F, Fiévet N, Barberger-Gateau P, Engelborghs S, De Deyn P, Mateo I, Franck A, Helisalmi S, Porcellini E, Hanon O, de Pancorbo MM, Lendon C, Dufouil C, Jaillard C, Leveillard T, Alvarez V, Bosco P, Mancuso M, Panza F, Nacmias B, Bossù P, Piccardi P, Annoni G, Seripa D, Galimberti D, Hannequin D, Licastrò F, Soininen H, Ritchie K, Blanché H, Dartigues JF, Tzourio C, Gut I, Van Broeckhoven C, Alperovitch A, Lathrop M, Amouyel P; European Alzheimer's Disease Initiative Investigators. Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. *Nat Genet.* 2009;41(10):1094-1099.
  7. Bertram L, Hiltunen M, Parkinson M, Ingelsson M, Lange C, Ramasamy K, Mullin K, Menon R, Sampson AJ, Hsiao MY, Elliott KJ, Velicelebi G, Moscarillo T, Hyman BT, Wagner SL, Becker KD, Blacker D, Tanzi RE. Family-based association between Alzheimer's disease and variants in *UBQLN1*. *N Engl J Med.* 2005;352(9):884-894.
  8. Blacker D, Albert MS, Bassett SS, Go RC, Harrell LE, Folstein MF; The National Institute of Mental Health Genetics Initiative. Reliability and validity of NINCDS-ADRDA criteria for Alzheimer's disease. *Arch Neurol.* 1994;51(12):1198-1204.
  9. McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology.* 1984;34(7):939-944.
  10. Brettschneider J, Petzold A, Schottle D, Claus A, Riepe M, Tumani H. The neurofilament heavy chain (NFH) in the cerebrospinal fluid diagnosis of Alzheimer's disease. *Dement Geriatr Cogn Disord.* 2006;21(5-6):291-295.
  11. Otto M, Wilfang J, Cepek L, Neumann M, Mollenhauer B, Steinacker P, Ciesielczyk B, Schulz-Schaeffer W, Kretzschmar HA, Poser S. Tau protein and 14-3-3 protein in the differential diagnosis of Creutzfeldt-Jakob disease. *Neurology.* 2002;58(2):192-197.
  12. Otto M, Esselmann H, Schulz-Schaeffer W, Neumann M, Schröter A, Ratzka P, Cepek L, Zerr I, Steinacker P, Windl O, Kornhuber J, Kretzschmar HA, Poser S, Wilfang J. Decreased beta-amyloid1-42 in cerebrospinal fluid of patients with Creutzfeldt-Jakob disease. *Neurology.* 2000;54(5):1099-1102.
  13. Reiber H, Thompson EJ, Grimsley G, Bernardi G, Adam P, Monteiro de Almeida S, Fredman P, Keir G, Lammers M, Liblau R, Menna-Barreto M, Sá MJ, Seres E, Sindic CJ, Teelken A, Trendelenburg C, Trojano M, van Antwerpen MP, Verbeek MM. Quality assurance for cerebrospinal fluid protein analysis: international consensus by an Internet-based group discussion. *Clin Chem Lab Med.* 2003;41(3):331-337.
  14. Lake SL, Blacker D, Laird NM. Family-based tests of association in the presence of linkage. *Am J Hum Genet.* 2000;67(6):1515-1525.
  15. Schjeide BM, McQueen MB, Mullin K, DiVito J, Hogan MF, Parkinson M, Hooli B, Lange C, Blacker D, Tanzi RE, Bertram L. Assessment of Alzheimer's disease case-control associations using family-based methods. *Neurogenetics.* 2009;10(1):19-25.
  16. Reiman EM, Webster JA, Myers AJ, Hardy J, Dunckley T, Zismann VL, Joshipura KD, Pearson JV, Hu-Lince D, Huentelman MJ, Craig DW, Coon KD, Liang WS, Herbert RH, Beach T, Rohrer KC, Zhao AS, Leung D, Bryden L, Marlowe L, Kaleem M, Mastroeni D, Grover A, Heward CB, Ravid R, Rogers J, Hutton ML, Melquist S, Petersen RC, Alexander GE, Caselli RJ, Kukull W, Papassotiropoulos A, Stephan DA. *GAB2* alleles modify Alzheimer's risk in *APOE* epsilon4 carriers. *Neuron.* 2007;54(5):713-720.
  17. Li H, Wetten S, Li L, St Jean PL, Upmanyu R, Surh L, Hosford D, Barnes MR, Briley JD, Borrie M, Coletta N, Delisle R, Dhalla D, Ehm MG, Feldman HH, Fornazari L, Gauthier S, Goodgame N, Guzman D, Hammond S, Hollingworth P, Hsiung GY, Johnson J, Kelly DD, Keren R, Kertesz A, King KS, Lovestone S, Loy-English I, Matthews PM, Owen MJ, Plumpton M, Pryse-Phillips W, Prinjha RK, Richardson JC, Saunders A, Slater AJ, St George-Hyslop PH, Stinnett SW, Swartz JE, Taylor RL, Wherrett J, Williams J, Yarnall DP, Gibson RA, Izratty MC, Middleton LT, Roses AD. Candidate single-nucleotide polymorphisms from a genome-wide association study of Alzheimer disease. *Arch Neurol.* 2008;65(1):45-53.
  18. Giedraitis V, Kilander L, Degerman-Gunnarsson M, Sundelöf J, Axelsson T, Sväven AC, Lannfelt L, Glaser A. Genetic analysis of Alzheimer's disease in the Uppsala Longitudinal Study of Adult Men. *Dement Geriatr Cogn Disord.* 2009;27(1):59-68.
  19. DerSimonian R, Laird N. Meta-analysis in clinical trials. *Control Clin Trials.* 1986;7(3):177-188.
  20. Harbord RM, Egger M, Sterne JA. A modified test for small-study effects in meta-analyses of controlled trials with binary endpoints. *Stat Med.* 2006;25(20):3443-3457.
  21. Ioannidis JP, Boffetta P, Little J, O'Brien TR, Uitterlinden AG, Vineis P, Balding DJ, Chokkalingam A, Dolan SM, Flanders WD, Higgins JP, McCarthy MI, McDermott DH, Page GP, Rebbeck TR, Seminara D, Khoury MJ. Assessment of cumulative evidence on genetic associations: interim guidelines. *Int J Epidemiol.* 2008;37(1):120-132.
  22. Khoury MJ, Bertram L, Boffetta P, Butterworth AS, Chanock SJ, Dolan SM, Fortier I, Garcia-Closas M, Gwinn M, Higgins JP, Janssens AC, Ostell J, Owen RP, Pagon RA, Rebbeck TR, Rothman N, Bernstein JL, Burton PR, Campbell H, Chokkalingam A, Furberg H, Little J, O'Brien TR, Seminara D, Vineis P, Winn DM, Yu W, Ioannidis JP. Genome-wide association studies, field synopses, and the development of the knowledge base on genetic variation and human diseases. *Am J Epidemiol.* 2009;170(3):269-279.
  23. Allen NC, Bagade S, McQueen MB, Ioannidis JP, Kavvoura FK, Khoury MJ, Tanzi RE, Bertram L. Systematic meta-analyses and field synopsis of genetic association studies in schizophrenia: the SzGene database. *Nat Genet.* 2008;40(7):827-834.
  24. Dubois B, Feldman HH, Jacova C, Dekosky ST, Barberger-Gateau P, Cummings J, Delacourte A, Galasko D, Gauthier S, Jicha G, Meguro K, O'Brien J, Pasquier F, Robert P, Rossor M, Salloway S, Stern Y, Visser PJ, Scheltens P. Research criteria for the diagnosis of Alzheimer's disease: revising the NINCDS-ADRDA criteria. *Lancet Neurol.* 2007;6(8):734-746.
  25. Kester MI, Blankenstein MA, Bouwman FH, van Elk EJ, Scheltens P, van der Flier WM. CSF biomarkers in Alzheimer's disease and controls: associations with APOE genotype are modified by age. *J Alzheimers Dis.* 2009;16(3):601-607.
  26. McCarthy MI, Abecasis GR, Cardon LR, Goldstein DB, Little J, Ioannidis JP, Hirschhorn JN. Genome-wide association studies for complex traits: consensus, uncertainty and challenges. *Nat Rev Genet.* 2008;9(5):356-369.
  27. Ioannidis JP, Thomas G, Daly MJ. Validating, augmenting and refining genome-wide association signals. *Nat Rev Genet.* 2009;10(5):318-329.
  28. Laird NM, Lange C. Family-based designs in the age of large-scale gene-association studies. *Nat Rev Genet.* 2006;7(5):385-394.
  29. Koo EH, Squazzo SL. Evidence that production and release of amyloid beta-protein involves the endocytic pathway. *J Biol Chem.* 1994;269(26):17386-17389.
  30. Dreyling MH, Martinez-Climent JA, Zheng M, Mao J, Rowley JD, Bohlander SK. The t(10;11)(p13;q14) in the U937 cell line results in the fusion of the *AF10* gene and *CALM*, encoding a new member of the AP-3 clathrin assembly protein family. *Proc Natl Acad Sci U S A.* 1996;93(10):4804-4809.
  31. Carey RM, Balcz BA, Lopez-Coviella I, Slack BE. Inhibition of dynamin-dependent endocytosis increases shedding of the amyloid precursor protein ectodomain and reduces generation of amyloid beta protein. *BMC Cell Biol.* 2005;6:30.