

Brain Monoamine Oxidase A Binding in Major Depressive Disorder

Relationship to Selective Serotonin Reuptake Inhibitor Treatment, Recovery, and Recurrence

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Context: Highly significant elevations in regional brain monoamine oxidase A (MAO-A) binding were recently reported during major depressive episodes (MDEs) of major depressive disorder (MDD). The relationship between MAO-A levels and selective serotonin reuptake inhibitor (SSRI) treatment, recovery, and recurrence in MDD is unknown.

Objectives: To determine whether brain MAO-A binding changes after SSRI treatment, whether brain MAO-A binding normalizes in subjects with MDD in recovery, and whether there is a relationship between prefrontal and anterior cingulate cortex MAO-A binding in recovery and subsequent recurrence of MDE.

Design: Case-control study.

Setting: Tertiary care psychiatric hospital.

Participants: Twenty-eight healthy subjects, 16 subjects with an MDE secondary to MDD, and 18 subjects with MDD in recovery underwent carbon 11–labeled harmine positron emission tomography scans. Subjects with MDE were scanned before and after 6 weeks of SSRI treatment. All were otherwise healthy, nonsmoking, and medication free. Subjects with MDD in recovery were followed up for 6 months after MAO-A binding measurement.

Main Outcome Measure: Monoamine oxidase A V_T , an index of MAO-A density, was measured in the prefrontal cortex, anterior cingulate cortex, posterior cingulate cortex, dorsal putamen, ventral striatum, thalamus, anterior temporal cortex, midbrain, and hippocampus.

Results: Monoamine oxidase A V_T was significantly elevated in each brain region both during MDE and after SSRI treatment as compared with healthy controls. During recovery, MAO-A V_T was significantly elevated in each brain region; however, those who went on to recurrence had significantly higher MAO-A V_T in the prefrontal and anterior cingulate cortex than those who did not.

Conclusions: Elevated MAO-A binding after SSRI treatment indicates persistence of a monoamine-lowering process not present in health. This provides a strong conceptual rationale for continuing SSRI treatment during early remission. Greater MAO-A binding in the prefrontal and anterior cingulate cortex in subjects with MDD in recovery and its association with subsequent recurrence argue that deficient monoamine neuro-modulation may persist into recovery and contribute to recurrence.

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MAJOR DEPRESSIVE DISORDER (MDD) has a high prevalence rate of 2% to 5% and is the fourth leading cause of death and disability.¹ It is increasingly recognized that resistance to treatment and recurrence of major depressive episodes (MDEs) constitute a large component of the burden of this illness.² The most common method to treat MDE and prevent subsequent recurrence is selective serotonin reuptake inhibitor (SSRI) treat-

ment. Given the burden of treatment resistance and recurrence, and the common use of SSRI, it is important to understand how neurobiological abnormalities in MDD are resistant to SSRI treatment and how neurobiological abnormalities persist into recovery.

An abnormality of highly significant elevations in monoamine oxidase A (MAO-A) binding in brain regions implicated in mood disorders was recently reported during MDE of MDD in a carbon 11 (¹¹C)–labeled harmine positron emis-

sion tomography (PET) study.³ [¹¹C]harmine is a selective, reversible PET radiotracer, with high brain uptake that binds with high affinity to MAO-A.⁴⁻⁷ Monoamine oxidase A is an enzyme found mainly on outer mitochondrial membranes that has an important role in the brain because it metabolizes serotonin, norepinephrine, and dopamine.^{8,9} Given the 2-SD effect size in MAO-A binding between subjects with MDE and healthy subjects in the earlier study, and the absence of other explanations to account for a primary monoamine-lowering process during MDE, elevated MAO-A density may be viewed as an important monoamine-lowering process during MDE.^{3,10-13} Moreover, Barton et al¹⁴ recently reported a consistent finding of elevated brain serotonin turnover in unmedicated depressed patients. Although MAO-A binding is elevated during an MDE, it is not clear whether elevated MAO-A binding persists after SSRI treatment and the relationship between MAO-A binding, recovery, and recurrence in MDD is unknown.

The first question that will be addressed in this study is does MAO-A binding normalize after SSRI treatment. While it is possible that many pathologies, such as elevated MAO-A binding, could normalize with remission of symptoms, we think the stronger argument is that SSRIs, by definition, do not target MAO-A directly. Therefore, the first main hypothesis is that MAO-A binding will remain persistently elevated after short-term (6 weeks) SSRI treatment in all brain regions, including those implicated in malfunctioning circuitry related to symptoms of MDEs (such as the prefrontal cortex, anterior cingulate cortex, thalamus, striatum, putamen, midbrain, hippocampus). The reason for expecting a persistent elevation in MAO-A binding in all brain regions rather than just a few regions is that MAO-A binding was elevated in every brain region assayed in the previous study of MAO-A binding during MDE.³ Identifying mismatches between SSRI treatment and neurobiology of MDD illness is important because these mismatches may be related to the serious clinical problems of not achieving remission in more than 40% of cases¹⁵ and high rates of relapse/recurrence during SSRI treatment that reach 20% over 2 years.¹⁶

The second question addressed in this study is does greater MAO-A binding occur during MDD in recovery. While monoamine dysregulation during MDD is most frequently investigated during MDE, there are reasons to suspect that the ability to regulate monoamines is impaired even in recovery. The most compelling evidence is that most studies report greater vulnerability to mood lowering in unmedicated subjects with MDD in recovery after tryptophan depletion and after dopamine and norepinephrine depletion.¹⁷⁻²³ Given these findings, the second main hypothesis of this study is that MAO-A binding will be elevated to a moderate extent in subjects with MDD in recovery.

Within this second question is an additional issue, namely, does greater MAO-A binding associate with subsequent recurrence of MDE? Greater MAO-A binding can be viewed as a monoamine-lowering process and it has been observed that chronically lowering monoamines, such as through reserpine administration, is associated with subsequent onset of MDEs.^{24,25} Therefore, an addi-

Table 1. Demographic and Clinical Characteristics Data of Healthy Subjects, Depressed Subjects, and Subjects With MDD in Recovery

| | Healthy (n=28) | Depressed (n=16) | MDD in Recovery (n=18) |
|---|----------------|------------------|------------------------|
| Age, y, mean (SD) | 31.57 (7.61) | 31.88 (8.39) | 31.11 (8.27) |
| HRSD score, mean (SD) | 0.61 (0.94) | 23.81 (2.97) | 1.17 (1.58) |
| Age at onset, y, mean (SD) | NA | 20.38 (8.97) | 19.28 (5.2) |
| No. of episodes, mean (SD) | 0 | 2.38 (1.36) | 3.65 (3.72) |
| Male, No. (%) | 15 (54) | 7 (44) | 8 (44) |
| Female, No. (%) | 13 (46) | 9 (56) | 10 (56) |
| Previous antidepressant treatment, ^a No. (%) | NA | 8 (50) | 12 (67) |
| Melancholic depression, No. (%) | NA | 2 (12.5) | 4 (22) |
| Family history of depression, No. (%) | 0 | 2 (12.5) | 7 (39) |

Abbreviations: HRSD, 17-item Hamilton Rating Scale for Depression; MDD, major depressive disorder; NA, not applicable.

^aDepressed subjects had no antidepressant treatment within the last 6 months and subjects with MDD in recovery had no antidepressant treatment within the last 12 months.

tional hypothesis is that recovered subjects with the greatest levels of MAO-A binding in the prefrontal and anterior cingulate cortex will have a greater risk of recurrence. These regions were chosen because their dysfunction is implicated in processes related to recurrence of MDEs. Both the prefrontal cortex and anterior cingulate cortex are often activated during mood-induction paradigms^{26,27} and during cognitive tests of negativistic perspective (such as anticipation of negative/positive events and loss aversion²⁸⁻³⁰).

METHODS

PARTICIPANTS

There were 3 groups of subjects: 28 healthy subjects (mean [SD] age, 31.6 [7.6] years, 15 men and 13 women), 16 subjects with MDE with MDD (mean [SD] age, 31.9 [8.4] years, 7 men and 9 women), and 18 subjects with MDD in recovery (mean [SD] age, 31.1 [8.3] years, 8 men and 10 women). Subjects with MDE were scanned before and after 6 weeks of SSRI treatment; of the 16 subjects with MDE, 15 returned for the second [¹¹C]harmine PET scan after treatment. All other subjects were scanned once. The sample of subjects with MDE is a different sample than that gathered for the earlier study by Meyer et al.³ Participants were within the age range of 19 to 49 years. Demographics for each group are listed in **Table 1**. For each study participant, written consent was obtained after the procedures had been fully explained. The study and recruitment procedures were approved by the Research Ethics Board for Human Subjects at the Centre for Addiction and Mental Health, University of Toronto.

All participants were physically healthy, did not smoke cigarettes, and had no history of neurotoxin or antipsychotic use. Participants were nonsmoking because cigarette smoking may lower MAO-A levels, which would create greater variability in MAO-A binding.³¹ Women in perimenopause or menopause were excluded. Healthy participants were screened to rule out

any Axis I disorders, and participants with MDD (with MDE and in recovery) were screened to rule out any comorbid Axis I disorders using the Structured Clinical Interview for DSM-IV.³² All participants were screened to rule out borderline and antisocial personality disorder using the Structured Clinical Interview for DSM-IV for Axis II disorders.³³ All subjects underwent common blood tests to rule out medical causes of disturbed mood (thyroid function, electrolyte levels, and complete blood cell count). All participants underwent a urine drug screen on the day of the [¹¹C]harmine PET scan. Use of over-the-counter medications and herbal remedies was exclusionary. Herbal remedies could not have been taken in the previous 3 months and over-the-counter medications/other medications could not have been taken in the previous month before scanning.

Subjects with current MDE were required to be antidepressant free for at least 6 months. No subject with current MDE had received antidepressant treatment within the past 7 months and 9 of the 16 patients with MDE had never received antidepressant treatment. Subjects with MDD in recovery were required to be antidepressant free for at least 1 year and not to have experienced an MDE for at least 1 year.

For all patients with MDD, a diagnosis of MDE secondary to MDD was based on the Structured Clinical Interview for DSM-IV for Axis I disorders³² and a consultation with a psychiatrist (J.H.M.). Additional exclusion criteria for all subjects with MDD included MDE with psychotic symptoms, bipolar disorder (type I or II), history of neuroleptic use, history of self-harm or suicidality outside episodes of depression, and history of alcohol or other drug abuse. For patients with MDE, the minimum severity for enrollment was based on a cutoff score of 20 on the 17-item Hamilton Rating Scale for Depression.³⁴ Subjects with MDE were treated with either citalopram hydrobromide with a titrating dose of 20 mg/d in the first week and 40 mg/d thereafter for 6 weeks or sertraline hydrochloride with a titrating dose of 50 mg/d in the first week and 100 mg/d thereafter for 6 weeks.

For subjects with MDD in recovery, a cutoff score of 7 or less on the 17-item Hamilton Rating Scale for Depression was required. Subjects with MDD in recovery were followed up for 6 months. They were assessed every 3 months (to cover the previous 3 months) with the depression module of the Structured Clinical Interview for DSM-IV for Axis I disorders³² and the Hamilton Rating Scale for Depression. In addition, subjects with MDD in recovery were encouraged to recontact the investigators (J.H.M., S.S., and L.M.) should symptoms recur. A diagnosis of a recurrent episode was verified by a psychiatrist (J.H.M.).

IMAGE ACQUISITION AND ANALYSIS

A dose of 370 MBq of intravenous [¹¹C]harmine was administered as a bolus for each PET scan. [¹¹C]harmine was of high radiochemical purity (mean [SD], 98.8% [0.7%]; n = 77) and high specific activity (mean [SD], 31.7 [18.2] TBq/mmol at the time of injection). An automatic blood sampling system was used to measure arterial blood radioactivity continuously for the first 10 minutes. The arterial input function was derived from the continuous whole-blood measurements from the automatic blood sampling system, the ratio of radioactivity in whole blood/plasma, and the percentage of unmetabolized radiotracer. The latter 2 measures were obtained from the manual samples, and a linear interpolation between the samples was applied to determine the whole-blood/plasma measurement in between the samples. Manual samples were obtained at 2.5, 7.5, 15, 20, 30, 45, 60, and approximately 90 minutes postinjection. The radioactivity in whole blood and plasma was measured as described previously.⁶

Frames were acquired as follows: 15 frames of 1 minute, then 15 frames of 5 minutes. The PET images were obtained using

a high-resolution research tomograph PET camera (in-plane resolution; full-width at half maximum, 3.1 mm; 207 axial sections of 1.2 mm; Siemens Molecular Imaging, Knoxville, Tennessee). Attenuation correction was done using a cesium 137-labeled transmission scan acquired in 64-bit list mode, which was converted into a 511-keV attenuation correction image.³⁵ Emission images were acquired in 64-bit list mode and were later reconstructed from 3-dimensional sinograms. Key steps in reconstruction included accounting for the octagonal design of the tomograph³⁶; correction for photon attenuation, detector normalization, and scatter in the 3-dimensional sinograms³⁷; Fourier rebinning to convert 3-dimensional into 2-dimensional sinograms³⁸; reconstruction into image space using a 2-dimensional filtered back-projection algorithm with a Hann filter at Nyquist cutoff frequency; and calibration of the images to nanocurie per cubic centimeter.

For the region of interest (ROI) method, each participant underwent magnetic resonance imaging (MRI) (GE Signa 1.5-T scanner; GE Medical Systems, Milwaukee, Wisconsin; fast spoiled gradient echo, T1-weighted image; x, y, and z voxel dimensions, 0.78, 0.78, and 1.5 mm). The ROIs were determined on MRIs that were coregistered to each summed [¹¹C]harmine PET image using a mutual information algorithm.³⁹ Regions of interest were determined using a semiautomated method in which regions on a template MRI were transformed onto the individual MRI via a series of transformation and deformation parameters that matched the template image to the coregistered MRI,^{40,41} followed by selection of gray matter voxels within the ROI.^{42,43} The location of the ROI was verified by visual assessment of the ROI on the coregistered MRI and summed [¹¹C]harmine PET image.

The ROIs selected were those for which abnormal function and/or neurochemistry has been implicated in mood regulation and/or mood disorders. The ROIs sampled the whole prefrontal cortex, anterior cingulate cortex (Brodmann areas 24 and part of 32), dorsal putamen, ventral striatum,⁴⁴ thalamus, anterior temporal cortex (Brodmann areas 38 and part of 20, 21, and 22), midbrain, and hippocampus.

The kinetics of [¹¹C]harmine can be described with an unconstrained 2-tissue compartment model.⁶ Highly identifiable fits with the unconstrained 2-tissue compartment model are obtainable for the V_T and may be readily determined for large data sets. The V_T is an index of harmine binding and represents the concentration of the total bound radiotracer in tissue relative to plasma concentration at equilibrium. The V_T can be expressed in terms of kinetic rate parameters as follows

$$V_T = (K_1/k_2) \times (k_3/k_4) + (K_1/k_2),$$

where K_1 and k_2 are influx and efflux rates for radiotracer passage across the blood-brain barrier and k_3 and k_4 describe the radioligand transfer between the free and nonspecific compartment and the specific binding compartment. K_1/k_2 is similar among different individuals (for further details, see Ginovart et al⁶). The [¹¹C]harmine PET measure of the MAO-A V_T was previously found to be reliable. Under test-retest conditions, applying the methods in this study on the high-resolution research tomograph scanner, the mean absolute difference in MAO-A V_T , expressed as a percentage of MAO-A V_T , ranged from 5% to 12% (n = 6 individuals) (J.H.M., A.A.W., S.H., et al, unpublished data, 2006).

STATISTICAL ANALYSIS

The primary analyses corresponded to the main hypotheses. A paired *t* test comparing MAO-A V_T before and after SSRI treat-

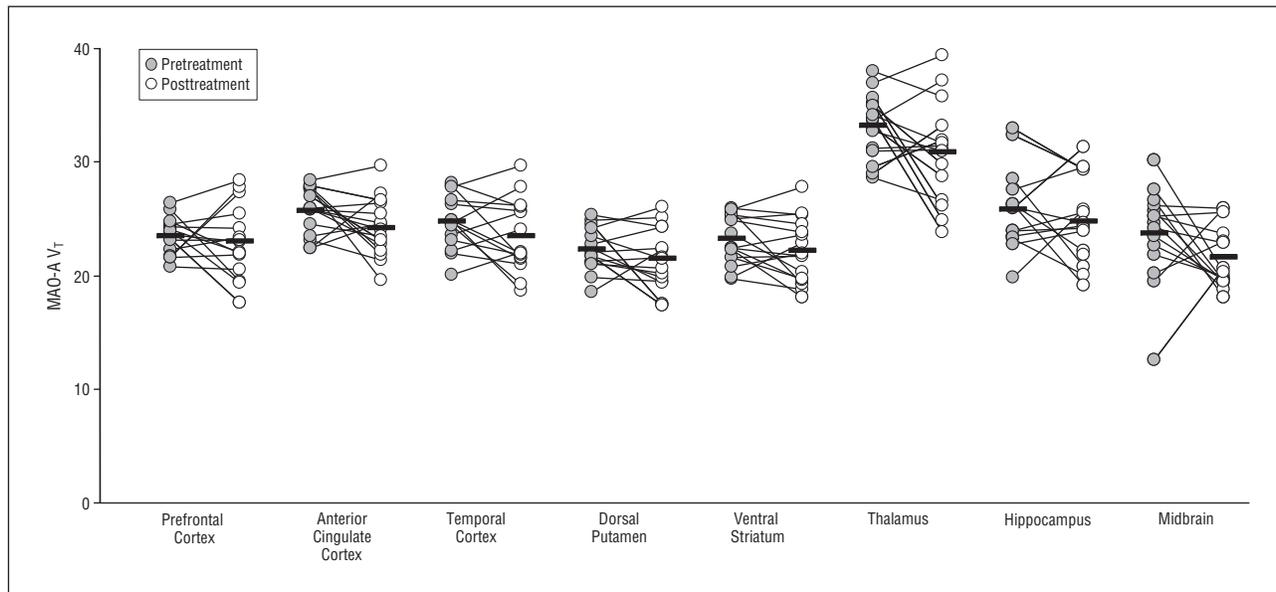


Figure 1. Regional monoamine oxidase A distribution volume (MAO-A V_T) before and after selective serotonin reuptake inhibitor treatment. Differences ranged from 1.6% to 9.2% of the original values and were nonsignificant (paired t test, $P=.68-.08$).

ment was applied in each region to assess treatment effect on MAO-A V_T . An independent-samples t test comparing MAO-A V_T between subjects with MDD in recovery and healthy individuals was applied to determine if MAO-A V_T differed between subjects with MDD in recovery and healthy individuals. Comparisons were done in each brain region for these analyses, since it was expected that MAO-A V_T would be affected in each brain region based on previous findings of elevated MAO-A binding during MDE.³ Finally, MAO-A V_T in the prefrontal cortex and anterior cingulate cortex were compared using an independent-samples t test between subjects with MDD in recovery who went on to have a recurrence and those subjects with MDD in recovery who did not go on to have a recurrence for the primary analysis, and then the same test was conducted for other regions for secondary analyses. It was planned not to include demographic variables in the analyses because it was expected that these would have no significant relationship with MAO-A binding.

RESULTS

As expected based on previous report, there were no effects of age or sex on levels of MAO-A V_T . Therefore, these variables were not included in subsequent analyses.

EFFECT OF SSRI TREATMENT ON MAO-A V_T IN SUBJECTS WITH CURRENT MDE

Within the subjects with a current MDE, the mean MAO-A V_T was similar before and after SSRI treatment in each region (paired t test, $t_{14}=0.42-1.88$; $P=.68-.08$). Post-treatment values were 1.6% to 9.2% less than pretreatment values (**Figure 1**). As would be expected in a sample of non-cigarette smoking, medication-naïve/medication-free, depressed subjects with no comorbid psychiatric or medical illness, a high proportion responded to treatment²: 12 subjects had a remission, 3 were nonresponders, and 1 was a partial responder. Fifteen subjects completed both [¹¹C]harmine PET scans. Post hoc

analysis found no relationship between SSRI type and change in MAO-A V_T in any region (repeated-measures analysis of variance, effect of medication type, $F_{1,13}=0.006-0.82$; $P=.94-.82$). Post hoc analysis showed no effect of lifetime antidepressant treatment exposure on baseline MAO-A V_T (independent-samples t test, $t_{13}=1.23-0.16$; $P=.24-.88$).

COMPARISON OF MAO-A V_T BETWEEN HEALTHY SUBJECTS, SUBJECTS WITH CURRENT MDE, AND SUBJECTS WITH MDD IN RECOVERY

The subjects with MDD in recovery had significantly higher MAO-A V_T than healthy subjects in each region (independent-samples t test, $t_{44}=2.3-4.9$; $P=.03$ to $P<.001$). Consistent with our previous report in a different sample,³ the subjects with a current MDE also had a significantly higher MAO-A V_T than healthy subjects in each region (independent-samples t test, $t_{42}=2.8-5.2$; $P=.007-.001$) (**Figure 2**). As expected, given the elevation in MAO-A V_T prior to treatment in the MDE group, and the lack of change in regional MAO-A V_T with treatment in the MDE group, regional MAO-A V_T was elevated in the SSRI-treated state as compared with healthy controls (independent-samples t test, $t_{41}=1.9-3.8$; $P=.06-.001$). Post hoc analysis showed no effect of lifetime antidepressant treatment exposure on baseline MAO-A V_T (independent-samples t test, $t_{13}=1.2-.16$; $P=.25-.88$).

ASSOCIATION BETWEEN MAO-A V_T AND RECURRENCE

The subjects with MDD in recovery who went on to have a recurrence had a significantly higher MAO-A V_T in the prefrontal cortex and anterior cingulate cortex than those subjects with MDD in recovery who did not go on to have a recurrence (independent-samples t test, prefrontal cor-

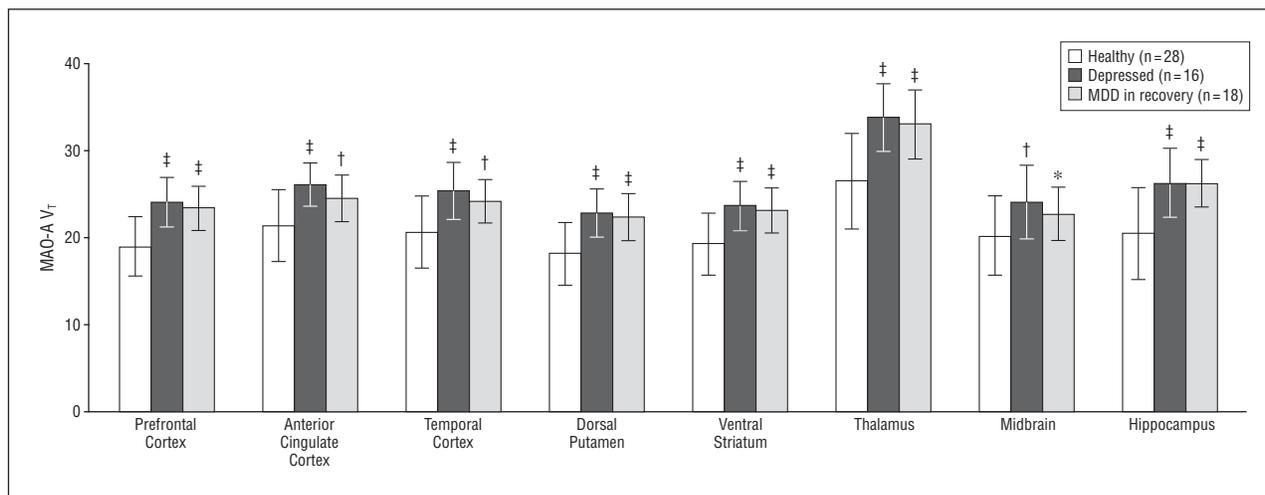


Figure 2. Monoamine oxidase A distribution volume (MAO-A V_T) in healthy subjects, depressed subjects, and subjects with major depressive disorder (MDD) in recovery. Monoamine oxidase A V_T was significantly greater in each patient sample as compared with healthy subjects (* $P < .05$, † $P < .01$, ‡ $P < .001$).

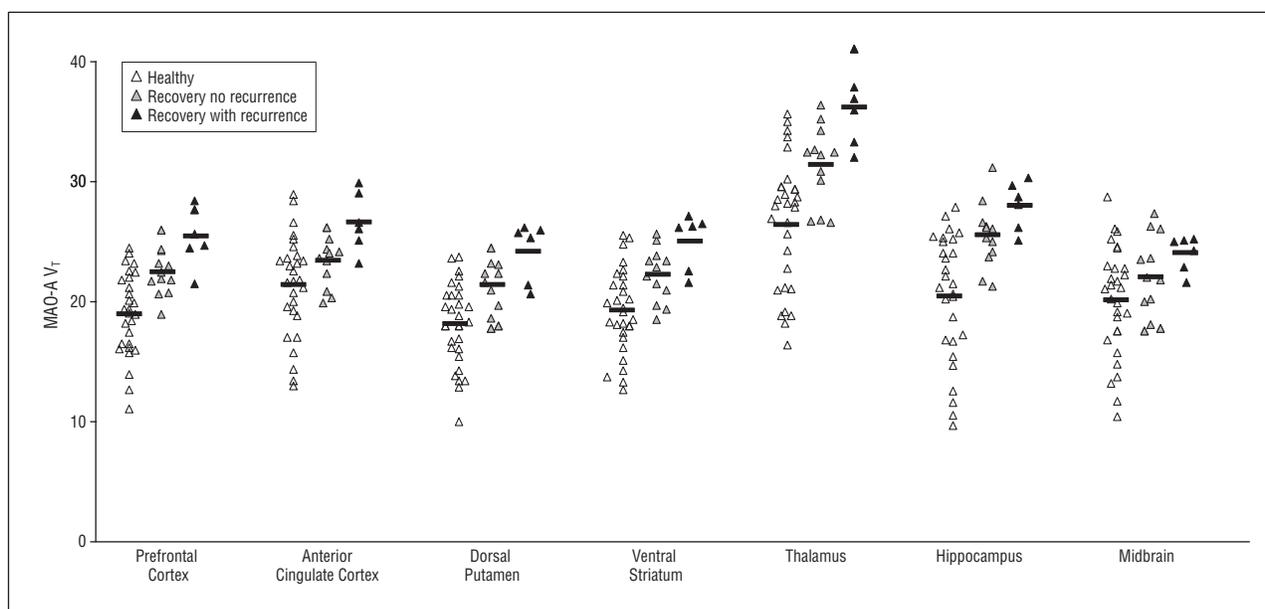


Figure 3. Monoamine oxidase A distribution volume (MAO-A V_T) in subjects with major depressive disorder (MDD) in recovery: relation to subsequent recurrence. Monoamine oxidase A V_T was significantly higher in subjects with MDD in recovery with recurrence as compared with other subjects with MDD in recovery in the prefrontal cortex ($P = .02$) and anterior cingulate cortex ($P = .02$) as well as the dorsal putamen, ventral striatum, and thalamus (all $P < .05$). Subjects with MDD in recovery with subsequent recurrence had significantly higher regional MAO-A V_T than healthy subjects in all brain regions ($P < .002$).

tex $t_{16} = 2.7$; $P = .02$; anterior cingulate cortex, $t_{16} = 2.7$; $P = .02$) (**Figure 3**). A similar pattern was observed in other regions (independent-samples t test, $t_{16} = 1.7-3.1$; $P = .11-.007$). There were no significant differences in the key demographic variables between those who went on to recurrence and those who did not as presented in **Table 2**.

COMMENT

The first main finding is that greater MAO-A binding during MDE persists after short-term SSRI treatment. This represents a partial mismatch between SSRI treatment and disease and provides important evidence to support a new model of serotonin dysregulation after SSRI treatment of

MDE in humans. The second main finding is that greater MAO-A binding occurs during recovery from MDD and is highest in those who have a recurrence in the subsequent 6 months. The second main finding argues for a scar or trait model of MAO-A binding elevation in MDD, supports a concept of ongoing monoamine deficit in relation to recurrence, and provides a new explanation for impaired monoamine regulation in MDD in recovery.

From the perspective of monoamine theory, SSRIs raise serotonin levels vigorously⁴⁵⁻⁴⁸ whereas elevated MAO-A levels would be expected to metabolize serotonin, norepinephrine, and dopamine excessively.⁹ The mismatch between monoamine levels raised by treatment and monoamine levels lowered by disease processes might, at times, contribute to lack of response to SSRI treatment. For ex-

Table 2. Demographic and Clinical Characteristics Data of Subjects With MDD in Recovery^a

| Characteristic | MDD in Recovery (n=18) | |
|---|------------------------|-----------------|
| | Recurrence | No Recurrence |
| Age, y, mean (SD) | 32.67 (11.76) | 30.33 (6.4) |
| HRSD score, mean (SD) | 1.17 (1.94) | 1.17 (1.47) |
| Age at onset, mean (SD) | 20.83 (7.55) | 18.42 (3.53) |
| No. of episodes, mean (SD) | 2.17 (1.47) | 2.42 (1.08) |
| Duration since last episode, wk, mean (SD) | 119.17 (173.34) | 124.83 (96.44) |
| Duration since last treatment, wk, mean (SD) | 125.00 (197.81) | 162.00 (175.50) |
| Male, No. (%) | 2 (33) | 6 (50) |
| Female, No. (%) | 4 (67) | 6 (50) |
| Previous antidepressant treatment, ^b No. (%) | 4 (67) | 6 (50) |
| Melancholic depression, No. (%) | 1 (17) | 3 (25) |
| Family history of depression, No. (%) | 3 (50) | 4 (33) |

Abbreviations: HRSD, 17-item Hamilton Rating Scale for Depression; MDD, major depressive disorder.

^aThere were no significant differences among demographic or clinical variables between groups.

^bNo antidepressant treatment within the last 12 months.

ample, motor slowing during MDE is associated with receptor binding changes consistent with striatal dopamine loss,^{11,13} and motor slowing during MDE was recently associated with lesser likelihood of response to fluoxetine hydrochloride.⁴⁹ From the perspective of the cellular vulnerability theory, MAO-A metabolism creates products that are potentially neurotoxic if present in excess, such as hydrogen peroxide, a reactive oxygen species.⁹ This may also reflect a mismatch between treatment and disease, since, to the best of our knowledge, it has not been demonstrated that inhibition of serotonin transporters directly influences production of reactive oxygen species.

Another important implication of persisting elevations in MAO-A levels in the SSRI-treated state is that the SSRI-treated state of an MDE is clearly not the same state as health, even when remission occurs. Greater levels of MAO-A in the SSRI-treated state can be incorporated into a new model of serotonin dysregulation during SSRI-treated MDE in humans (**Figure 4**). Key elements in this model include excess MAO-A, an 80% SSRI blockade at the serotonin transporter, and a modest reduction in binding of some serotonin receptors. It is well established that SSRIs block 80% of serotonin transporter sites in humans at doses that are clinically superior to placebo.^{50,51} In medication-free subjects with MDE, a modest reduction in cortex serotonin 2A (5-hydroxytryptamine 2A [5-HT_{2A}]) binding potential after SSRI treatment has been reported⁴⁸ and subjects with MDE who have had SSRI treatment within the past month often have reductions in 5-HT_{2A} binding.⁵²⁻⁵⁶

This model has strong clinical relevance. It is standard practice to maintain SSRI treatment beyond the initial 6-week trial when remission is present. This is supported by the clinical finding that cessation of antidepressant use after short-term treatment is associ-

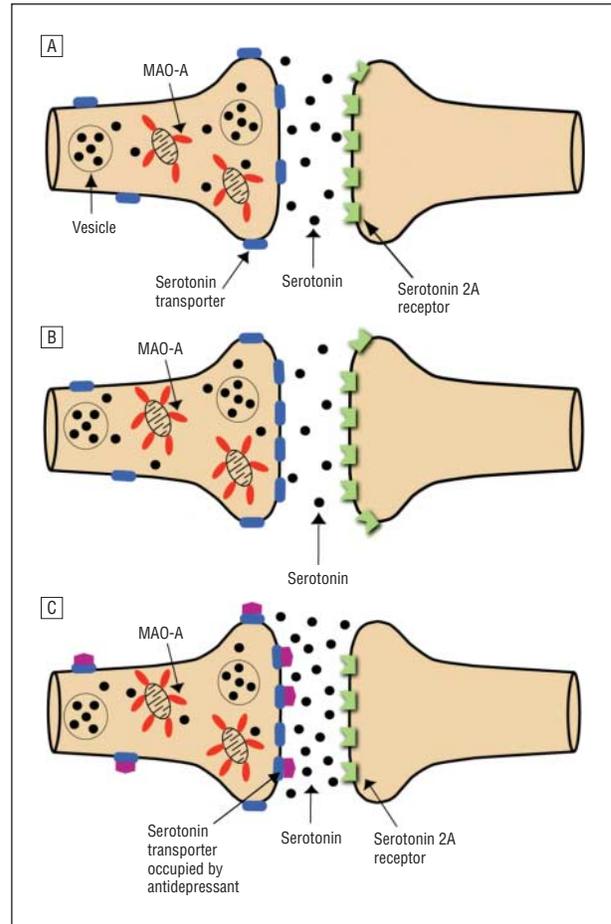


Figure 4. Monoamine theory for selective serotonin reuptake inhibitor (SSRI)-treated major depressive episode (MDE). A, Health. B, The state of an untreated MDE with greater monoamine oxidase A (MAO-A) binding and some degree of excessive serotonin transporter binding. C, The SSRI-treated MDE with persistently elevated MAO-A binding, an 80% serotonin transporter blockade, and a modestly reduced concentration of serotonin 2A receptors.^{48,50-52}

ated with a 40% relapse rate over the subsequent 6 months whereas continuation of antidepressant treatment over the same period is associated with an 18% relapse rate.^{57,58} A notable implication of this model of SSRI-treated MDE is that stopping SSRI use after short-term treatment is problematic because a key monoamine-lowering process is still present. This provides a new theoretical argument to explain why it is extremely important to continue SSRI treatment to prevent relapse following the initial 6-week trial.

Greater MAO-A levels provide a mechanistic explanation for some abnormalities of receptor binding and monoamine regulation in MDD in recovery. Greater 5-HT_{2A} binding was recently reported in medication-free subjects with MDD in recovery who had strong histories for recurrent MDD.⁵⁹ The finding of greater MAO-A binding in recovery in the present study is consistent with this result. Greater MAO-A levels would be expected to excessively metabolize 5-HT and there is an inverse relationship between long-term manipulations of serotonin levels and cortex 5-HT_{2A} binding.⁶⁰⁻⁶³ Investigations of tryptophan depletion often report more frequent mood lowering in unmedicated subjects with MDD in recov-

ery as compared with healthy subjects.^{17,19,21,64,65} Greater MAO-A levels may explain greater vulnerability to tryptophan depletion in MDD in recovery through excess metabolism of 5-HT by MAO-A, which would facilitate loss of extracellular 5-HT. Similarly, greater MAO-A binding in MDD in recovery could explain the particular vulnerability to mood lowering after depletion of dopamine and norepinephrine after α -methylparatyrosine administration^{22,23} since MAO-A participates in the removal of both monoamines. Thus, greater MAO-A levels in recovery may be considered a process contributing to impaired control of monoamines.

The relationship between greater MAO-A binding in the prefrontal and anterior cingulate cortex with subsequent recurrence argues that monoamine-lowering processes contribute to recurrence of MDE. This may appear to be inconsistent with some views of the monoamine theory of MDD that suggest that low monoamine levels must occur simultaneously with reductions in mood or presence of symptoms. However, it has been reported that long periods of monoamine depletion, such as after reserpine administration, can lead to MDE in humans.^{24,25,66} Even so, the reports that long-term administration of a monoamine-lowering medication can lead to MDE are not equivalent to the idea that monoamine-lowering processes actually occur in unmedicated patients with MDD in recovery. The present study argues a new point that monoamine-lowering processes may occur during MDD in recovery and, when more prominent, are associated with recurrence.

Since greater brain MAO-A binding occurs during recovery and during MDE, greater MAO-A binding may be considered a scar or a trait. Future work will need to differentiate between scar and trait processes and identify the underlying mechanism for elevations in MAO-A binding during recovery. Etiologies to consider include both genetic, epigenetic, and hormonal influences.^{9,67,68} Unfortunately, the current intervention of SSRI treatment is not directly intervening on MAO-A binding. In the longer-term, it may be therapeutic to target the underlying processes that raise MAO-A levels in recovery so as to better prevent future MDE.

There are limitations of measurement and interpretation in this study. By using a neuroimaging ligand technique, we were able to measure MAO-A binding in vivo but the resolution of the technique does not clarify whether the binding is increased within a cell or organelle (like mitochondria membrane) or whether more MAO-A-containing cells or organelles are present. The measure of MAO-A used, an index of MAO-A density called MAO-A V_T , reflects total binding, has the advantage of being computationally efficient, and is the most stable and least variable measure of [¹¹C]harmine binding. However, approximately 15% of this measure reflects free and nonspecific binding so it is assumed that free and nonspecific binding do not differ tremendously between groups.⁶ An elevation in MAO-A V_T may also reflect greater affinity of MAO-A, although this would not change our interpretation as greater affinity of MAO-A for monoamines would be expected to contribute to monoamine loss. A limitation of the monoamine-oriented interpretations of the MAO-A binding abnor-

mality is that the MAO-A binding abnormality may also be related to other abnormalities in MDD. For example, changes in MAO-A binding could be elevated secondary to other processes, such as mitochondrial dysfunction,⁶⁹ that then participate in the pathophysiology of MDD.

There are also some limitations in the statistical design. The first issue is analyzing multiple data sets. There are 3 main data sets in this study: the MAO-A binding before and after SSRI treatment, the MAO-A binding in health and in subjects with MDD in recovery, and the MAO-A binding data in the subjects with MDD in recovery combined with the follow-up data for recurrence. As more data sets are analyzed in a laboratory, the likelihood of a chance finding rises. For example, after completion of 5 separate studies, each with a main significance threshold of $P = .05$, for which the true result should be nonsignificant in each, there would be a 25% likelihood of at least 1 significant finding due to chance alone. The second issue is that we examined multiple regions. For the main analyses of investigating how MAO-A binding changes after SSRI treatment, and the comparison of MAO-A binding between subjects with MDD in recovery and healthy subjects, all regions were included in the hypothesis. For the relationship between MAO-A binding and recurrence, the prefrontal cortex and anterior cingulate cortex regions were included in the hypothesis. For all of these findings, the expected results occurred in each of the hypothesized regions. Should it have been that only 1 region significantly met the hypothesis, then the finding could have been due to chance alone as a result of multiple comparisons. For example, in a study of 10 hypothesized regions, in which 1 region alone is significant at a P value of .05, there would be a 50% likelihood that the single regional finding was due to chance alone.

To our knowledge, this is the first study to measure brain MAO-A binding before and after SSRI treatment and brain MAO-A binding in unmedicated subjects with MDD in recovery. Monoamine oxidase A binding in all regions remained consistently elevated before and after SSRI treatment, even in subjects in remission, indicating a persistent abnormality that is inadequately targeted by SSRI treatment. It also demonstrates that the SSRI-treated state, even when symptoms are in remission, is clearly different from health and provides theoretical support for why discontinuation of SSRI use after short-term treatment is associated with a high risk of relapse. In subjects with MDD in recovery, regional MAO-A binding was also elevated, and greater levels of MAO-A binding in the prefrontal and anterior cingulate cortex were associated with recurrence. This argues that monoamine-lowering processes may occur during MDD in recovery and are associated with recurrence. Elevated MAO-A binding in recovery also provides an explanation for the frequently observed mood lowering after monoamine depletions in this group¹⁷⁻²³ and identifies an important treatment target not previously implicated as a pathophysiological abnormality in the recovery phase of MDD.

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REFERENCES

1. Ustün TB, Ayuso-Mateos JL, Chatterji S, Mathers C, Murray CJ. Global burden of depressive disorders in the year 2000. *Br J Psychiatry*. 2004;184:386-392.
2. Trivedi MH, Rush AJ, Wisniewski SR, Nierenberg AA, Warden D, Ritz L, Norquist G, Howland RH, Lebowitz B, McGrath PJ, Shores-Wilson K, Biggs MM, Balasubramani GK, Fava M; STAR*D Study Team. Evaluation of outcomes with citalopram for depression using measurement-based care in STAR*D: implications for clinical practice. *Am J Psychiatry*. 2006;163(1):28-40.
3. Meyer JH, Ginovart N, Boovariwala A, Sagrati S, Hussey D, Garcia A, Young T, Praschak-Rieder N, Wilson AA, Houle S. Elevated monoamine oxidase A levels in the brain: an explanation for the monoamine imbalance of major depression. *Arch Gen Psychiatry*. 2006;63(11):1209-1216.
4. Bergström M, Westerberg G, Kihlberg T, Langstrom B. Synthesis of some 11C-labelled MAO-A inhibitors and their in vivo uptake kinetics in rhesus monkey brain. *Nucl Med Biol*. 1997;24(5):381-388.
5. Bergström M, Westerberg G, Langstrom B. 11C-harmine as a tracer for monoamine oxidase A (MAO-A): in vitro and in vivo studies. *Nucl Med Biol*. 1997;24(4):287-293.
6. Ginovart N, Meyer JH, Boovariwala A, Hussey D, Rabiner EA, Houle S, Wilson AA. Positron emission tomography quantification of [11C]-harmine binding to monoamine oxidase-A in the human brain. *J Cereb Blood Flow Metab*. 2006;26(3):330-344.
7. Tweedie DJ, Burke MD. Metabolism of the beta-carbolines, harmine and harmol, by liver microsomes from phenobarbitone- or 3-methylcholanthrene-treated mice: identification and quantitation of two novel harmine metabolites. *Drug Metab Dispos*. 1987;15(1):74-81.
8. Haefely W, Burkard WP, Cesura AM, Kettler R, Lorez HP, Martin JR, Richards JG, Scherschlicht R, Da Prada M. Biochemistry and pharmacology of moclobemide, a prototype RIMA. *Psychopharmacology (Berl)*. 1992;106(suppl):S6-S14.
9. Youdim MB, Edmondson D, Tipton KF. The therapeutic potential of monoamine oxidase inhibitors. *Nat Rev Neurosci*. 2006;7(4):295-309.
10. Meyer JH, Houle S, Sagrati S, Carella A, Hussey DF, Ginovart N, Goulding V, Kennedy J, Wilson AA. Brain serotonin transporter binding potential measured with carbon 11-labeled DASB positron emission tomography: effects of major depressive episodes and severity of dysfunctional attitudes. *Arch Gen Psychiatry*. 2004;61(12):1271-1279.
11. Meyer JH, Kruger S, Wilson AA, Christensen BK, Goulding VS, Schaffer A, Minié C, Houle S, Hussey D, Kennedy SH. Lower dopamine transporter binding potential in striatum during depression. *Neuroreport*. 2001;12(18):4121-4125.
12. Meyer JH, McMain S, Kennedy SH, Korman L, Brown GM, DaSilva JN, Wilson AA, Blak T, Eynan-Harvey R, Goulding VS, Houle S, Links P. Dysfunctional attitudes and 5-HT(2) receptors during depression and self-harm. *Am J Psychiatry*. 2003;160(1):90-99.
13. Meyer JH, McNeely HE, Sagrati S, Boovariwala A, Martin K, Verhoeff NP, Wilson AA, Houle S. Elevated putamen D(2) receptor binding potential in major depression with motor retardation: an [11C]raclopride positron emission tomography study. *Am J Psychiatry*. 2006;163(9):1594-1602.
14. Barton DA, Esler MD, Dawood T, Lambert EA, Haikerwal D, Brenchley C, Socrates F, Hastings J, Guo L, Wiesner G, Kaye DM, Bayles R, Schlaich MP, Lambert GW. Elevated brain serotonin turnover in patients with depression: effect of genotype and therapy. *Arch Gen Psychiatry*. 2008;65(1):38-46.
15. Thase ME. Evaluating antidepressant therapies: remission as the optimal outcome. *J Clin Psychiatry*. 2003;64(suppl 13):18-25.
16. Frank E, Kupfer DJ, Perel JM, Cornes C, Jarrett DB, Mallinger AG, Thase ME, McEachran AB, Grochocinski VJ. Three-year outcomes for maintenance therapies in recurrent depression. *Arch Gen Psychiatry*. 1990;47(12):1093-1099.
17. Leyton M, Young SN, Blier P, Ellenbogen MA, Palmour RM, Ghadirian AM, Benkelfat C. The effect of tryptophan depletion on mood in medication-free, former patients with major affective disorder. *Neuropsychopharmacology*. 1997;16(4):294-297.
18. Smith KA, Clifford EM, Hockney RA, Clark DM, Cowen PJ. Effect of tryptophan depletion on mood in male and female volunteers: a pilot study. *Hum Psychopharmacol*. 1997;12(2):111-117.
19. Moreno FA, Gelenberg AJ, Heninger GR, Potter RL, McKnight KM, Allen J, Phillips AP, Delgado PL. Tryptophan depletion and depressive vulnerability. *Biol Psychiatry*. 1999;46(4):498-505.
20. Moreno FA, Heninger GR, McGahuey CA, Delgado PL. Tryptophan depletion and risk of depression relapse: a prospective study of tryptophan depletion as a potential predictor of depressive episodes. *Biol Psychiatry*. 2000;48(4):327-329.
21. Neumeister A, Nugent AC, Waldeck T, Geraci M, Schwarz M, Bonne O, Bain EE, Luckenbaugh DA, Herscovitch P, Charney DS, Drevets WC. Neural and behavioral responses to tryptophan depletion in unmedicated patients with remitted major depressive disorder and controls. *Arch Gen Psychiatry*. 2004;61(8):765-773.
22. Berman RM, Narasimhan M, Miller HL, Anand A, Cappiello A, Oren DA, Heninger GR, Charney DS. Transient depressive relapse induced by catecholamine depletion: potential phenotypic vulnerability marker? *Arch Gen Psychiatry*. 1999;56(5):395-403.
23. Hasler G, Fromm S, Carlson PJ, Luckenbaugh DA, Waldeck T, Geraci M, Roiser JP, Neumeister A, Meyers N, Charney DS, Drevets WC. Neural response to catecholamine depletion in unmedicated subjects with major depressive disorder in remission and healthy subjects. *Arch Gen Psychiatry*. 2008;65(5):521-531.
24. Freis ED. Mental depression in hypertensive patients treated for long periods with large doses of reserpine. *N Engl J Med*. 1954;251(25):1006-1008.
25. Burrell RH. Depression associated with reserpine therapy. *NZ Med J*. 1956;55(307):228-231.
26. Liotti M, Mayberg HS, McGinnis S, Brannan SL, Jerabek P. Unmasking disease-specific cerebral blood flow abnormalities: mood challenge in patients with remitted unipolar depression. *Am J Psychiatry*. 2002;159(11):1830-1840.
27. Krüger S, Alda M, Young LT, Goldapple K, Parikh S, Mayberg HS. Risk and resilience markers in bipolar disorder: brain responses to emotional challenge in bipolar patients and their healthy siblings. *Am J Psychiatry*. 2006;163(2):257-264.
28. Knutson B, Taylor J, Kaufman M, Peterson R, Glover G. Distributed neural representation of expected value. *J Neurosci*. 2005;25(19):4806-4812.
29. Tom SM, Fox CR, Trepel C, Poldrack RA. The neural basis of loss aversion in decision-making under risk. *Science*. 2007;315(5811):515-518.
30. Sharot T, Riccardi AM, Raio CM, Phelps EA. Neural mechanisms mediating optimism bias. *Nature*. 2007;450(7166):102-105.
31. Fowler JS, Volkow ND, Wang GJ, Pappas N, Logan J, Shea C, Alexoff D, MacGregor RR, Schlyer DJ, Zezulova I, Wolf AP. Brain monoamine oxidase A inhibition in cigarette smokers. *Proc Natl Acad Sci U S A*. 1996;93(24):14065-14069.
32. First M, Spitzer R, Williams J, Gibbon M. *Structured Clinical Interview for DSM-IV Axis I Disorders, Patient Edition (SCID-P), version 2*. New York, NY: Biometrics Research; 1995.
33. Blais MA, Norman DK. A psychometric evaluation of the DSM-IV personality disorder criteria. *J Pers Disord*. 1997;11(2):168-176.
34. Hamilton M. A rating scale for depression. *J Neurol Neurosurg Psychiatry*. 1960;23:56-62.
35. Knoess C, Rist J, Michel C, Burbar Z, Eriksson L, Panin V, Byars L, Lenox M, Wienhard K, Heiss W-D, Nutt R. Evaluation of single photon transmission for the HRRT. *IEEE Nucl Sci Symp Conf Rec*. 2003;3:1936-1940.
36. Karp JS, Muehlechner G, Lewitt R. Constrained Fourier space method for compensation of missing data in emission computed tomography. *IEEE Trans Med Imaging*. 1988;7(1):21-25.
37. Watson C, Newport D, Casey M, deKemp R, Beanlands R, Schmand M. Evaluation of simulation-base scatter correction for 3D PET cardiac imaging. *IEEE Trans Nucl Sci*. 1996;44:90-97.
38. Defrise M, Kinahan P, Townsend D, Michel C, Sibomana M, Newport D. Exact and approximate rebinning algorithms for 3-D PET data. *IEEE Trans Med Imaging*. 1997;16(2):145-158.

39. Studholme C, Hill D, Hawkes D. An overlap invariant entropy measure of 3D medical image alignment. *Pattern Recognit.* 1999;32(1):71-86.
40. Ashburner J, Friston KJ. Nonlinear spatial normalization using basis functions. *Hum Brain Mapp.* 1999;7(4):254-266.
41. Ashburner J, Neelin P, Collins DL, Evans A, Friston K. Incorporating prior knowledge into image registration. *Neuroimage.* 1997;6(4):344-352.
42. Rusjan P, Mamo D, Ginovart N, Hussey D, Vitcu I, Yasuno F, Tetsuya S, Houle S, Kapur S. An automated method for the extraction of regional data from PET images. *Psychiatry Res.* 2006;147(1):79-89.
43. Ashburner J, Friston K. Multimodal image coregistration and partitioning—a unified framework. *Neuroimage.* 1997;6(3):209-217.
44. Mawlawi O, Martinez D, Slifstein M, Broft A, Chatterjee R, Hwang DR, Huang Y, Simpson N, Ngo K, Van Heertum R, Laruelle M. Imaging human mesolimbic dopamine transmission with positron emission tomography, I: accuracy and precision of D(2) receptor parameter measurements in ventral striatum. *J Cereb Blood Flow Metab.* 2001;21(9):1034-1057.
45. Bel N, Artigas F. Chronic treatment with fluvoxamine increases extracellular serotonin in frontal cortex but not in raphe nuclei. *Synapse.* 1993;15(3):243-245.
46. Tatsumi M, Groshan K, Blakely RD, Richelson E. Pharmacological profile of antidepressants and related compounds at human monoamine transporters. *Eur J Pharmacol.* 1997;340(2-3):249-258.
47. Owens MJ, Morgan WN, Plott SJ, Nemeroff CB. Neurotransmitter receptor and transporter binding profile of antidepressants and their metabolites. *J Pharmacol Exp Ther.* 1997;283(3):1305-1322.
48. Meyer JH, Kapur S, Eisefeld B, Brown GM, Houle S, DaSilva J, Wilson AA, Rafi-Tari S, Mayberg HS, Kennedy SH. The effect of paroxetine upon 5-HT_{2A} receptors in depression: an [^{18F}]setoperone PET imaging study. *Am J Psychiatry.* 2001;158(1):78-85.
49. Taylor BP, Bruder GE, Stewart JW, McGrath PJ, Halperin J, Ehrlichman H, Quitkin FM. Psychomotor slowing as a predictor of fluoxetine nonresponse in depressed outpatients. *Am J Psychiatry.* 2006;163(1):73-78.
50. Meyer JH, Wilson AA, Ginovart N, Goulding V, Hussey D, Hood K, Houle S. Occupancy of serotonin transporters by paroxetine and citalopram during treatment of depression: a [(11C)]DASB PET imaging study. *Am J Psychiatry.* 2001;158(11):1843-1849.
51. Meyer JH, Wilson AA, Sagrati S, Hussey D, Carella A, Potter WZ, Ginovart N, Spencer EP, Cheok A, Houle S. Serotonin transporter occupancy of five selective serotonin reuptake inhibitors at different doses: an [(11C)]DASB positron emission tomography study. *Am J Psychiatry.* 2004;161(5):826-835.
52. Meyer JH. Applying neuroimaging ligands to study major depressive disorder. *Semin Nucl Med.* 2008;38(4):287-304.
53. Biver F, Wikler D, Lotstra F, Damhaut P, Goldman S, Mendlewicz J. Serotonin 5-HT₂ receptor imaging in major depression: focal changes in orbito-insular cortex. *Br J Psychiatry.* 1997;171:444-448.
54. Mintun MA, Sheline YI, Moerlein SM, Vlassenko AG, Huang Y, Snyder AZ. Decreased hippocampal 5-HT_{2A} receptor binding in major depressive disorder: in vivo measurement with [^{18F}]altanserin positron emission tomography. *Biol Psychiatry.* 2004;55(3):217-224.
55. Yatham LN, Liddle PF, Shiah IS, Scarrow G, Lam RW, Adam MJ, Zis AP, Ruth TJ. Brain serotonin₂ receptors in major depression: a positron emission tomography study. *Arch Gen Psychiatry.* 2000;57(9):850-858.
56. Stockmeier CA. Involvement of serotonin in depression: evidence from post-mortem and imaging studies of serotonin receptors and the serotonin transporter. *J Psychiatr Res.* 2003;37(5):357-373.
57. Geddes JR, Carney SM, Davies C, Furukawa TA, Kupfer DJ, Frank E, Goodwin GM. Relapse prevention with antidepressant drug treatment in depressive disorders: a systematic review. *Lancet.* 2003;361(9358):653-661.
58. Simons AD, Murphy GE, Levine JL, Wetzel RD. Cognitive therapy and pharmacotherapy for depression: sustained improvement over one year. *Arch Gen Psychiatry.* 1986;43(1):43-48.
59. Bhagwagar Z, Hinz R, Taylor M, Fancy S, Cowen P, Grasby P. Increased 5-HT (2A) receptor binding in euthymic, medication-free patients recovered from depression: a positron emission study with [(11C)]MDL 100,907. *Am J Psychiatry.* 2006;163(9):1580-1587.
60. Stockmeier CA, Kellar KJ. In vivo regulation of the serotonin-2 receptor in rat brain. *Life Sci.* 1986;38(2):117-127.
61. Roth BL, McLean S, Zhu X, Chuang D. Characterization of two [3H]ketanserin recognition sites in rat striatum. *J Neurochem.* 1987;49(6):1833-1838.
62. Todd KG, McManus DJ, Baker GB. Chronic administration of the antidepressants phenelzine, desipramine, clomipramine, or maprotiline decreases binding to 5-hydroxytryptamine_{2A} receptors without affecting benzodiazepine binding sites in rat brain. *Cell Mol Neurobiol.* 1995;15(3):361-370.
63. O'Regan D, Kwok RP, Yu PH, Bailey BA, Greenshaw AJ, Boulton AA. A behavioural and neurochemical analysis of chronic and selective monoamine oxidase inhibition. *Psychopharmacology (Berl).* 1987;92(1):42-47.
64. Smith KA, Fairburn CG, Cowen PJ. Relapse of depression after rapid depletion of tryptophan. *Lancet.* 1997;349(9056):915-919.
65. Ruhé HG, Mason NS, Schene AH. Mood is indirectly related to serotonin, norepinephrine and dopamine levels in humans: a meta-analysis of monoamine depletion studies. *Mol Psychiatry.* 2007;12(4):331-359.
66. Mendels J, Frazer A. Brain biogenic amine depletion and mood. *Arch Gen Psychiatry.* 1974;30(4):447-451.
67. Pinsonneault JK, Papp AC, Sadee W. Allelic mRNA expression of X-linked monoamine oxidase a (MAOA) in human brain: dissection of epigenetic and genetic factors. *Hum Mol Genet.* 2006;15(17):2636-2649.
68. Shih JC, Chen K, Ridd MJ. Monoamine oxidase: from genes to behaviour. *Annu Rev Neurosci.* 1999;22:197-217.
69. Shao L, Martin MV, Watson SJ, Schatzberg A, Akil H, Myers RM, Jones EG, Bunney WE, Vawter MP. Mitochondrial involvement in psychiatric disorders. *Ann Med.* 2008;40(4):281-295.